

Synthesis of Teleocidins A, B and Their Congeners. Part 2.¹ Synthesis of Lyngbyatoxin A (Teleocidin A-1), Teleocidin A-2, Pendolmycin, and (*R, E*)- and (*S, E*)-7-(3,7,11-Trimethyl-1,6,10- dodecatrien-3-yl)-(-)-indolactams V

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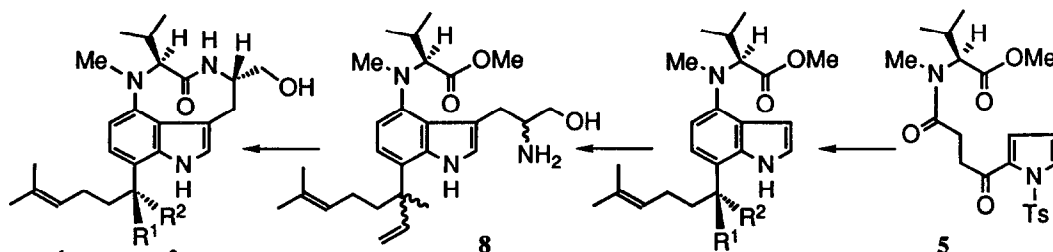
(Received in Japan 30 July 1991)

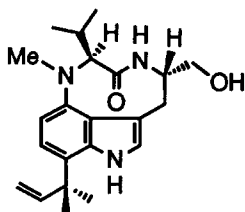
Key Words Alkaloid synthesis, Lyngbyatoxin A, Teleocidin A-2, Pendolmycin, Teleocidin analogue

Abstract—Details of the synthesis method of the tumor promoters, lyngbyatoxin A (= teleocidin A-1) (1) and teleocidin A-2 (2) from (*R*)- and (*S*)-methyl *N*-[7-(3,7-dimethyl-1,6-octadien-3-yl)-4-indolyl]-*N*-methyl-L-valinate (3 and 4) are presented. Other titled compounds, 6, 7a, and 7b, were prepared analogously.

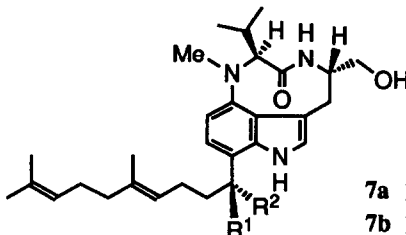
In the previous paper we reported the details of preparation of (*R*)- and (*S*)-methyl *N*-[7-(3,7-dimethyl-1,6-octadien-3-yl)-4-indolyl]-*N*-methyl-L-valinates (3 and 4) starting from the readily available methyl *N*-methyl-L-valinate derivative 5. Here we describe continuation of our synthesis to obtain lyngbyatoxin A (= teleocidin A-1) (1) and teleocidin A-2 (2). This synthesis pathway was also applied to the preparation of pendolmycin (6) and (*R, E*)- and (*S, E*)-7-(3,7,11-trimethyl-1,6,10-dodecatrien-3-yl)-(-)-indolactams V (7a and 7b) to send them for biological evaluation.

With the essential intermediates 3 and 4 in hand, the next task was to introduce the amino propanol side chain at the C-3 position of the indole moiety to form 8 and to set the stage to make the nine-membered lactam ring to complete the natural product synthesis. This was first tried by applying conventionally either the Vilsmeier reagent or the gramine formation (Chart 1). Reaction of 9a with the Vilsmeier reagent gave the expected product 10a in 65% yield, but the formation of a position isomer 11a was unavoidable, in 22% yield. On the other hand, treating 9a, 9b, and an inseparable mixture¹ of 9c and 9d (9c + 9d) with *N, N*-dimethyl(methylene)ammonium chloride² in dichloromethane at room temperature afforded only the expected compounds 12a, 12b, and 12c + 12d in 97%, 95%, and 88% yields, respectively. And the gramine compounds





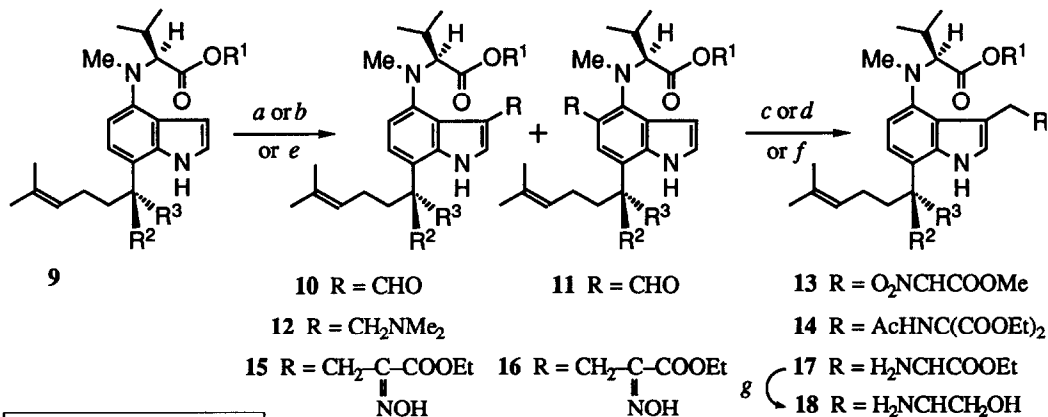
6 Pendolmycin

7a R¹ = vinyl R² = Me7b R¹ = Me R² = vinyl

12b and 12c + 12d were transformed into a nitrodiester 13b and acetamide triesters 14c + 14d in 91% and 49% yields, respectively, by a reaction with methyl nitroacetate³ and diethyl acetamidomalonate with the aid of tributylphosphine⁴ However, modest yield of 14c + 14d precluded subsequent experiments for trying decarboxylation Reduction of the nitro group of 13b with aluminium-amalgam⁵ or the cobaltous chloride – sodium borohydride reagent⁶ was unsuccessful

A satisfactory result was obtained using ethyl 3-bromo-2-hydroxyiminopropanoate.⁷ The respective reaction with 9a and 9b was effected in the presence of sodium carbonate in dichloromethane at room temperature to afford 15a and 15b in 59% and 65% yields, accompanied by the formation of regioisomers 16a and 16b in 20% and 15% yields Reduction of the oxime function to the primary amine group was readily carried out by treatment of 15a and 15b with aluminium – amalgam in aqueous tetrahydrofuran⁸ to furnish 17a and 17b in 92% and 89% yields Judging from their proton NMR spectra, each was composed of a 1:1 mixture of diastereomers with respect to the amino group configuration, and therefore no stereochemical control was unfortunately operated during the reduction The chemical reactivity of the newly formed amino ester group was different from the *N*-methylvalinate ester, probably due to difference in the environmental situation Thus the reduction of 17a and 17b using sodium borohydride in refluxing ethanol⁹ afforded 18a and 18b in 51% and 53% yields

The final step, the formation of a nine-membered lactam ring, was performed as reported by Ley and co-workers for indolactam V synthesis¹⁰ The amino ester 18a was hydrolyzed at reflux for 22 h with 10% potassium hydroxide in a 4:1 mixture of methanol and water (Chart 2) The potassium salt of the amino acid was



| | R ¹ | R ² | R ³ |
|---|----------------|----------------|----------------|
| a | Me | vinyl | Me |
| b | Me | Me | vinyl |
| c | <i>t</i> -Bu | vinyl | Me |
| d | <i>t</i> -Bu | Me | vinyl |

a Vilsmeier reagent b Me₂N⁺=CH₂ Cl⁻ c O₂NCH₂COOMe
 d AcHNCH(COOEt)₂, Bu₃P e BrCH₂-C(=NOH)-COOEt, Na₂CO₃
 f Al-Hg, THF-H₂O g NaBH₄, EtOH

Chart 1

neutralized with triethylamine hydrochloride, and after removing the solvent and thoroughly drying the remaining solid, the residue was treated with diphenylphosphoryl azide (DPPA)¹¹ and triethylamine in dimethylformamide at room temperature for 17 h. The reaction products were separated and purified by the usual silica gel chromatography and subsequent HPLC to afford lyngbyatoxin A (= teleocidin A-1) (**1**), the unnatural enantiomer of teleocidin A-2 (ent-**2**), and an inseparable mixture of 9-epilyngbyatoxin A (9-epi-**1**) and 9-epi-ent-**2** in 23%, *ca* 4%, and 27% yields, respectively. The same series of treatments of **18b** afforded analogously teleocidin A-2 (**2**), the enantiomer of lyngbyatoxin A (ent-**1**), and an inseparable mixture of 9-epiteleocidin A-2 (9-epi-**2**) and 9-epi-ent-**1** in 21%, *ca* 4%, and 25% yields, respectively. The two final compounds **1** and **2** as well as two by-products ent-**1** and ent-**2** obtained here were converted to the (+)-MTPA [(+)- α -methoxy- α -trifluoromethylphenylacetic acid] esters and their pure states were confirmed by HPLC analysis. The synthetic compounds **1** and **2** were identified as lyngbyatoxin A (= teleocidin A-1) and teleocidin A-2 by comparing their MS, ¹H and ¹³C NMR, and CD spectra, and the mobility on HPLC. They also exhibited the same order of biological activities as the natural products on the tests of mouse ear irritation, induction of ornithine decarboxylase, inhibition of specific ³H-TPA binding, and activation of protein kinase C.

The structures of ent-**1** and ent-**2** were supported by the antipodal CD curves, together with the identical MS, ¹H NMR spectra, and the mobility on HPLC, compared with those of **1** and **2**. The origin of these enantiomers ent-**1** and ent-**2** as well as their C-9 epimers 9-epi-ent-**1** and 9-epi-ent-**2** was attributable to a rather drastic treatment with alkali during hydrolysis of the methyl ester group in **18a** and **18b**. Partial racemization took place at the L-valine portion and the combination of this inversion (12*S* \rightarrow 12*R*) with other chiralities at the C-9 and C-14 positions, for instance (9*S*, 14*R*), generated the compound 9-epi-ent-**2**. This phenomenon was inevitable when the methyl valine ester was used, since it resisted the other mild hydrolysis conditions.

Thus the first synthesis of lyngbyatoxin A (teleocidin A-1) (**1**) and teleocidin A-2 (**2**) was achieved in

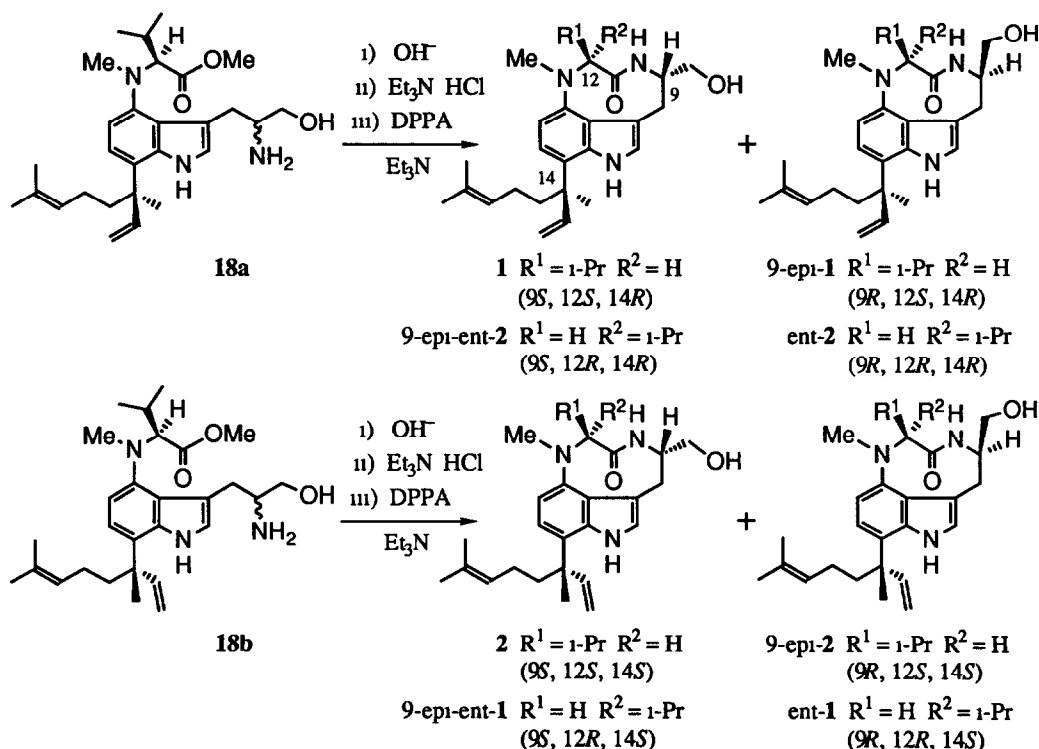


Chart 2

eleven steps from 1-[(4-methylphenyl)sulfonyl]pyrrole. The success of this straightforward and relatively short synthesis was due to the pertinent selection of the starting intermediate **5**, which made it possible to introduce the linalyl side chain in a single step and to efficiently produce the 7-alkyl-4-aminoindole derivatives **3** and **4**. Introduction of the aminopropanol side chain was readily achieved and subsequent formation of the nine-membered ring afforded the final compounds, although some unwanted by-products were unavoidably produced.

This synthesis pathway is widely applicable to other teleocidins and related compounds. So next we attempted to synthesize pendolmycin (**6**), a metabolite of *Nocardioopsis* strain SA 1715, and (*R, E*)- and (*S, E*)-7-(3,7,11-trimethyl-1,6,10-dodecatrien-3-yl)-(-)-indolactams **V** (**7a** and **7b**), unnatural analogues of lyngbyatoxin A (**1**) (Chart 3). The common starting intermediate was the ketoamide **5** and the Grignard reaction of **5** with 3-methyl-2-butenyl (prenyl) bromide or (*E, E*)-3,7,11-trimethyl-2,6,10-dodecatrienyl (farnesyl) bromide¹² in the presence of magnesium afforded mixtures of **19a** and **20a**, and **19b** and **20b**. These were separated and the *N*-tosyl group was reductively cleaved from **19a** and **19b** with magnesium in methanol. Combined pyrrole derivatives **20a** and **20b** were obtained in 84% and 66% yields, respectively. Both were dehydrated without trouble in 93% and 91% yields to afford **21a** and **21b**, whose amide function was changed to the thioamide group with Lawesson's reagent giving **22a** and **22b** in 74% and 77% yields. The indole cyclization was carried out as before by treating these thioamides with iodomethane in dimethylformamide to yield **23a** (66%), **23b** (46.5%), and **24b** (25%), accompanied by the formation of by-products **25a** (19%) and **25b** (23%).

The carbon side chain was introduced into the C-3 position of the indole derivatives **23a**, **23b** and **24b** as usual by reacting ethyl 3-bromo-2-hydroxyiminopropanoate. The desired compounds **26a**, **26b** and **27b** were produced in 53%, 51% and 61% yields together with by-products **28a** ($R^2 = \text{Me}$, $R^3 = \text{vinyl}$) (21%), **28b** ($R^2 = \text{vinyl}$, $R^3 = \text{Me}$) (20%), **28b** ($R^2 = \text{Me}$, $R^3 = \text{vinyl}$) (14%), **29a** ($R^2 = \text{Me}$, $R^3 = \text{vinyl}$) (4%), **29b** ($R^2 = \text{vinyl}$, $R^3 = \text{Me}$) (4%), **29b** ($R^2 = \text{Me}$, $R^3 = \text{vinyl}$) (3%), and diastereoisomeric mixtures of **30a** ($R^2 = \text{Me}$, $R^3 = \text{vinyl}$) (4%), **30b** ($R^2 = \text{vinyl}$, $R^3 = \text{Me}$) (2.5%), and **30b** ($R^2 = \text{Me}$, $R^3 = \text{vinyl}$) (3%). The oxime group in **26a**, **26b** and **27b** was reduced with aluminium-amalgam to give aminodiester compounds **31a**, **31b** and **32b** in 92%, 95% and 94% yields and their unhindered ester group was reduced with sodium borohydride in the presence of lithium chloride,¹³ followed by conversion to the trifluoroacetates. Diastereoisomers related to the amino group stereochemistry were separated here, affording sets of epimers **33a** (40%) and **34a** (37.5%), **33b** (40%) and **34b** (33%), as well as **35b** (38.5%) and **36b** (33%). Among them, the compounds produced in the better yields, *i.e.* **33a**, **33b** and **35b**, afforded the required products, pendolmycin (**37a** = **6**), and (*R, E*)- and (*S, E*)-7-(3,7,11-trimethyl-1,6,10-dodecatrien-3-yl)-(-)-indolactams **V** (**37b** = **7a** and **39b** = **7b**) in 54%, 56% and 47% yields by a series of operations: i) alkaline hydrolysis of the methyl valinate part, ii) neutralization of the alkaline medium, iii) complete dryness of the reaction residue, and iv) treatment with diethylphosphoryl cyanide (DEPC)¹⁴ (see Part 3, the following paper) and triethylamine in dimethylformamide. Partial racemization during the alkaline hydrolysis gave rise to the formation of **38a**, **38b** and **40b** in 9%, 12% and 9% yields.

The tumor promoting activities of pendolmycin (**6**) and 7-[(*R, E*)-3,7,11-trimethyl-1,6,10-dodecatrien-3-yl]-(-)-indolactam **V** (**7a**) were studied using our specimens and it was shown that the activity of **7a** was as potent as that of teleocidins A, whereas **6** was moderately active and its potency was somewhere between teleocidins A and (-)-indolactam **V**.¹⁵

EXPERIMENTAL

General Procedures — Mass spectra (MS) were recorded on Hitachi RMS-4 and Hitachi M-80B spectrometers. High resolution mass spectra (HRMS) were measured on JEOL JMS-DX-300 and Hitachi M-80B spectrometers. Circular dichroism spectra (CD) were taken on JASCO J-500A. Proton magnetic resonance (¹H NMR) spectra were recorded on Varian EM 390 (90 MHz), JEOL FX-200A (270 MHz) and JEOL JNM-GX-400 (400 MHz) spectrometers in CDCl₃ with TMS as an internal reference. Preparative thin-layer chromatography over Al₂O₃ (Al₂O₃ PTLC) was carried out on glass plates (20 × 20 cm) coated with Merck aluminum oxide 60 PF₂₅₄ (type E) (1 mm thick). Other General Methods were described in the Part 1.¹

Vilsmeier Reaction on 9a to Form 10a and 11a — Vilsmeier-Haak reagent (0.5 ml, 0.098 mmol),

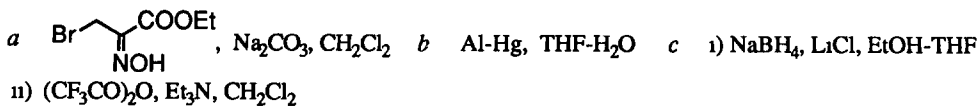
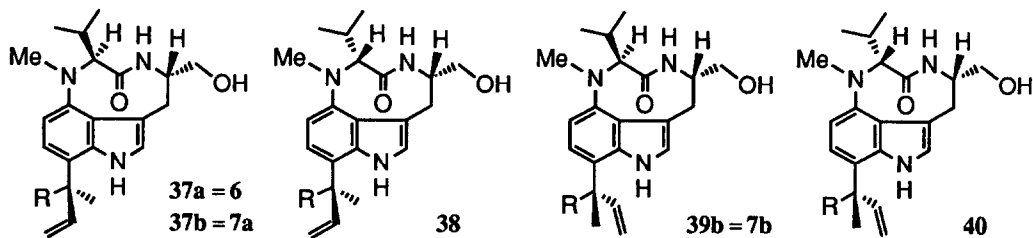
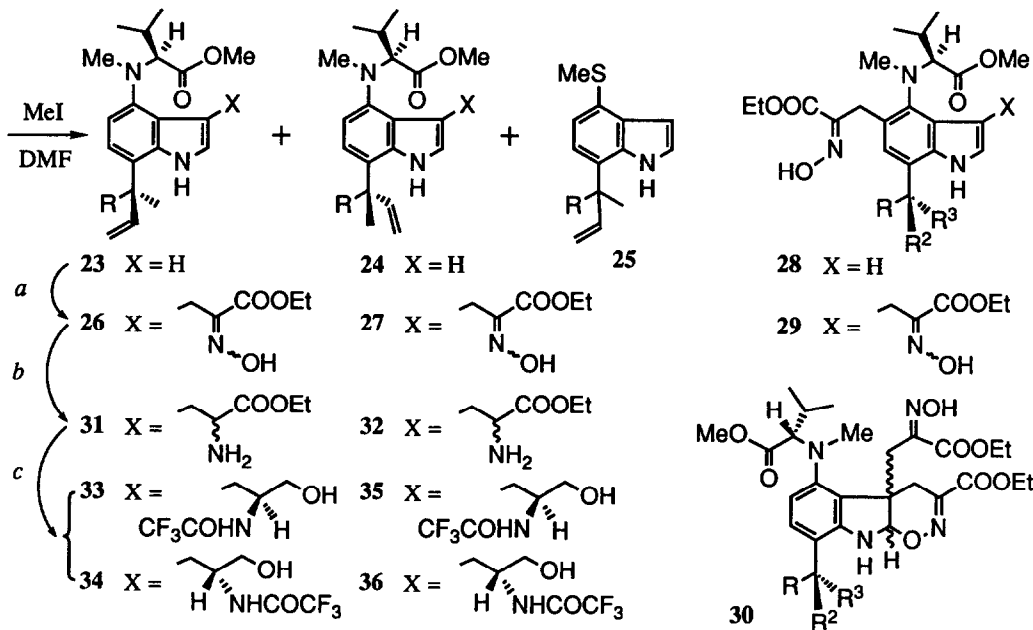
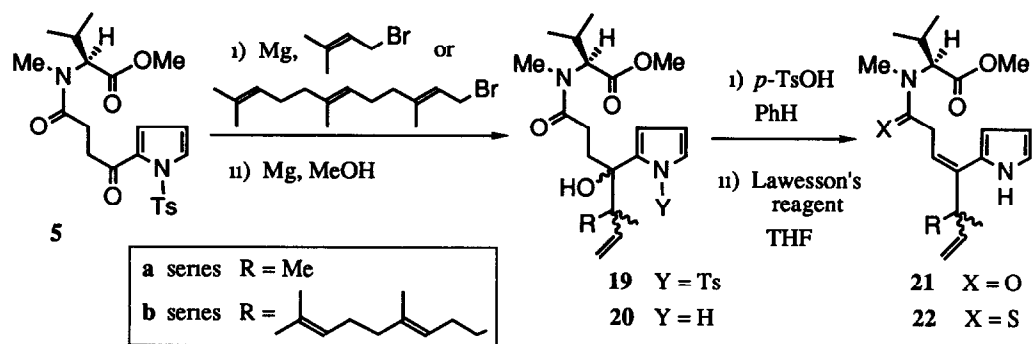


Chart 3

prepared from POCl_3 (60 mg, 0.391 mmol) and DMF (2 ml) with stirring at 0°C for 5 min, was added to a solution of **9a** (13 mg, 0.033 mmol) in Et_2O (1 ml), and the mixture was stirred at 0°C for 30 min. Sat. $\text{NaHCO}_3\text{-H}_2\text{O}$ (4 ml) was added to this and the whole was stirred vigorously at room temperature for 20 min. Extraction with Et_2O , usual work-up and PTLC [hexane-EtOAc (4:1)] gave **10a** (9 mg, 65%) and **11a** (3 mg, 22%). **Methyl *N*-[3-Formyl-7-[(*R*)-3,7-dimethyl-1,6-octadien-3-yl]-4-indolyl]-*N*-methyl-L-valinate (10a)** Colorless syrup MS m/z 424 (M^+) IR (CHCl_3) cm^{-1} 1728, 1653 $^1\text{H NMR}$ (90 MHz) δ 0.90 (3H, d, $J=6.5$ Hz), 1.09 (3H, d, $J=6.5$ Hz), 1.41 (3H, s), 1.47 (3H, s), 1.61 (3H, s), 2.92 (3H, s), 3.55 (3H, s), 3.66 (1H, d, $J=9$ Hz), 4.89-5.16 (1H, m), 5.29 (1H, dd, $J=18, 1.5$ Hz), 5.32 (1H, dd, $J=10.5, 1.5$ Hz), 6.24 (1H, dd, $J=18, 10.5$ Hz), 7.08 (2H, s), 7.91 (1H, d, $J=3$ Hz, changed to s with D_2O), 9.25 (1H, br s, NH), 10.60 (1H, s) **Methyl *N*-[5-Formyl-7-[(*R*)-3,7-dimethyl-1,6-octadien-3-yl]-4-indolyl]-*N*-methyl-L-valinate (11a)** Colorless syrup MS m/z 424 (M^+) IR (CHCl_3) cm^{-1} 1723, 1658 $^1\text{H NMR}$ (90 MHz) δ 0.90 (3H, d, $J=6.5$ Hz), 1.18 (3H, d, $J=6.5$ Hz), 1.41 (3H, s), 1.50 (3H, s), 1.61 (3H, s), 3.22 (3H, s), 3.44 (3H, s), 3.59 (1H, d, $J=9.5$ Hz), 4.88-5.14 (1H, m), 5.25 (1H, dd, $J=18, 1.5$ Hz), 5.27 (1H, dd, $J=10, 1.5$ Hz), 6.19 (1H, dd, $J=18, 10.5$ Hz), 6.65 (1H, dd, $J=3, 2$ Hz), 7.09 (1H, dd, $J=3, 2.5$ Hz), 7.64 (1H, s), 8.78 (1H, br s, NH), 10.50 (1H, s)

Reaction of 9a, 9b and 9c + 9d with *N,N*-Dimethyl(methylene)ammonium Chloride to Form Gramine Derivatives 12a, 12b and 12c + 12d — Preparation of **12a** is typical. A solution of **9a** (37 mg, 0.093 mmol) in CH_2Cl_2 (2 ml) was stirred with *N,N*-dimethyl(methylene)ammonium chloride (25 mg, 0.267 mmol) at room temperature for 1 h 20 min. Sat. $\text{NaHCO}_3\text{-H}_2\text{O}$ was added, the whole was extracted with CH_2Cl_2 , and the extract was worked up as usual. Purification by Al_2O_3 PTLC [hexane-EtOAc (3:2)] afforded **methyl *N*-[3-(dimethylamino)methyl-7-[(*R*)-3,7-dimethyl-1,6-octadien-3-yl]-4-indolyl]-*N*-methyl-L-valinate (12a)** (41 mg, 97%) as colorless syrup MS m/z 453 (M^+) IR (CHCl_3) cm^{-1} 1727, 1630 $^1\text{H NMR}$ (90 MHz) δ 0.90 (3H, d, $J=6.5$ Hz), 1.13 (3H, d, $J=6.5$ Hz), 1.38 (3H, s), 1.44 (3H, s), 1.60 (3H, s), 2.34 (6H, s), 2.86 (3H, s), 3.47 (3H, s), 3.58 (1H, d, $J=8.5$ Hz), 3.85 (1H, d, $J=13.5$ Hz), 4.12 (1H, d, $J=13.5$ Hz), 4.90-5.18 (1H, m), 5.23 (1H, dd, $J=10.5, 1.5$ Hz), 5.24 (1H, dd, $J=18, 1.5$ Hz), 6.21 (1H, dd, $J=18, 10.5$ Hz), 6.80 (1H, d, $J=8$ Hz), 6.95 (1H, d, $J=8$ Hz), 7.08 (1H, d, $J=2.5$ Hz, changed to s with D_2O), 8.53 (1, br s, NH) **Methyl *N*-[3-(Dimethylamino)methyl-7-[(*S*)-3,7-dimethyl-1,6-octadien-3-yl]-4-indolyl]-*N*-methyl-L-valinate (12b)** Colorless syrup MS m/z 453 (M^+) IR (CHCl_3) cm^{-1} 1727, 1628 $^1\text{H NMR}$ (90 MHz) δ 0.90 (3H, d, $J=6.5$ Hz), 1.10 (3H, d, $J=6.5$ Hz), 1.38 (3H, s), 1.43 (3H, s), 1.60 (3H, s), 2.30 (6H, s), 2.83 (3H, s), 3.52 (3H, s), 3.60 (1H, d, $J=8.5$ Hz), 3.83 (1H, d, $J=13.5$ Hz), 4.09 (1H, d, $J=13.5$ Hz), 4.89-5.19 (1H, m), 5.22 (1H, dd, $J=10.5, 1.5$ Hz), 5.23 (1H, dd, $J=18, 1.5$ Hz), 6.20 (1H, dd, $J=18, 10.5$ Hz), 6.80 (1H, d, $J=8$ Hz), 6.95 (1H, d, $J=8$ Hz), 7.06 (1H, d, $J=2$ Hz, changed to s with D_2O), 8.50 (1, br s, NH) ***t*-Butyl *N*-[3-(Dimethylamino)methyl-7-(3,7-dimethyl-1,6-octadien-3-yl)-4-indolyl]-*N*-methyl-L-valinate (12c + 12d)** Colorless amorphous compound MS m/z 495 (M^+) IR (CHCl_3) cm^{-1} 1715 $^1\text{H NMR}$ (90 MHz) of two diastereomers δ 1.24 (9H, s), 1.43 (6H, s), 1.62 (3H, br s), 2.36 (6H, s), 2.83 and 2.80 (3H, s each), 3.51 and 3.54 (1H, d each, $J=10.5$ Hz), 3.94 (1H, d, 1.21 Hz), 4.13 (1H, d, $J=14$ Hz), 4.94-5.18 (1H, m), 5.24 (1H, dd, $J=10.5, 2.5$ Hz), 5.26 (1H, dd, $J=18, 2.5$ Hz), 6.21 (1H, dd, $J=18, 10.5$ Hz), 6.89 (1H, d, $J=8$ Hz), 6.99 (1H, d, $J=8$ Hz), 7.12 (1H, br d, $J=2$ Hz, changed to s with D_2O), 8.53 (1, br s, NH)

Methyl *N*-[7-[(*S*)-3,7-Dimethyl-1,6-octadien-3-yl]-3-(2-methoxycarbonyl-2-nitro)ethyl-4-indolyl]-*N*-methyl-L-valinate (13b) — A solution of **12b** (18 mg, 0.040 mmol) and methyl nitroacetate (40 mg, 0.336 mmol) in toluene (2 ml) was refluxed for 3 h under Ar atmosphere. After evaporation to dryness, the residue was purified by PTLC [hexane-EtOAc (4:1)] to afford **13b** (19 mg, 91%), colorless syrup MS m/z 527 (M^+) IR (CHCl_3) cm^{-1} 1752, 1730, 1630, 1559, 1373 $^1\text{H NMR}$ (90 MHz) of major and minor diastereomers (*ca.* 3:2) δ 0.91 and 0.88 (3H, d each, $J=6.5$ Hz), 1.04 and 0.99 (3H, d each, $J=6.5$ Hz), 1.33 and 1.38 (3H, s each), 1.42 (3H, s), 1.60 (3H, s), 2.73 and 2.76 (3H, s each), 3.41 and 3.48 (3H, s each), 3.79 and 3.67 (3H, s each), 4.88-5.17 (1H, m), *ca.* 5.90-6.32 (1H, m), 6.18 (1H, dd, $J=18, 10.5$ Hz), 8.49 (1H, br s, NH)

***t*-Butyl *N*-[3-(2-Acetamido-2,2-diethoxycarbonyl)ethyl-7-(3,7-dimethyl-1,6-octadien-3-yl)-4-indolyl]-*N*-methyl-L-valinate (14c + 14d)** — A solution of **12c + 12d** (6 mg, 0.012 mmol), diethyl acetamidomaloate (8 mg, 0.037 mmol) and 95% *n*- Bu_3P (5 mg, 0.024 mmol) in CH_3CN (2 ml) was refluxed under N_2 atmosphere for 1.5 h. After evaporation to dryness, the residue was purified by PTLC (3% MeOH- CH_2Cl_2) to give a crude mixture (10 mg), which was further purified by PTLC [benzene-EtOAc (5:1)] to furnish **14c + 14d** (4 mg, 49%), colorless syrup MS m/z 667 (M^+) IR (CHCl_3) cm^{-1} 1730, 1680 $^1\text{H NMR}$ (90 MHz, 60°C) δ 1.21 (9H, s), 1.41 (6H, s), 1.61 (3H, s), 1.88 (3H, s), 2.07-2.49 (1H, m), 2.90 (3H, s), 3.49 (1H, d, $J=8$ Hz), 3.86-4.34 (6H, m), 4.92-5.12 (1H, m), 6.17 (1H, dd, $J=18, 10.5$ Hz), 6.83-7.04 (4H, m), 8.40 (1H, br s, NH)

Reaction of 9a and 9b with Ethyl 3-Bromo-2-hydroxyiminopropanoate to Form 15a, 16a, 15b and

16b — Preparation of **15a** and **16a** is typical. To a solution of **9a** (124 mg, 0.313 mmol) in CH_2Cl_2 (6 ml) were added successively Na_2CO_3 (66 mg, 0.623 mmol) and ethyl 3-bromo-2-hydroxyiminopropanoate (66 mg, 0.314 mmol) under Ar atmosphere and the mixture was stirred at 15°C for 15 h. The mixture was filtered through a celite bed and the celite was rinsed with CH_2Cl_2 . The combined organic layer was washed with sat. $\text{NH}_4\text{Cl-H}_2\text{O}$ and worked up as usual. Purification by PTLC (0.5% EtOH- CH_2Cl_2) afforded the recovered **9a** (10 mg, 8%), **16a** (33 mg, 20%) and the crude **15a** (99 mg) in the order of increasing polarity. The crude **15a** was purified by PTLC [hexane-EtOAc (3:1)] to give ethyl 7-[(*R*)-3,7-dimethyl-1,6-octadien-3-yl]- α -(hydroxyimino)-4-[[(*S*)-1-(methoxycarbonyl)-2-methylpropyl]methylamino]indole-3-propanoate (**15a**) (97 mg, 59%) as colorless syrup. HRMS Calcd for $\text{C}_{30}\text{H}_{43}\text{N}_3\text{O}_5$ 525.3202. Found: 525.3222. IR (CHCl_3) cm^{-1} 1722, 1628. $^1\text{H NMR}$ (90 MHz) δ 0.91 (3H, d, $J=6.5$ Hz), 1.11 (3H each, d, $J=6.5$ Hz), 1.18 (3H, t, $J=7$ Hz), 1.40 (6H, s), 1.60 (3H, s), 2.91 (3H, s), 3.51 (3H, s), 3.66 (1H, d, $J=9$ Hz), 4.21 (2H, q, $J=7$ Hz), 4.36 (1H, d, 15.5 Hz), 4.56 (1H, d, $J=15.5$ Hz), 4.89-5.20 (1H, m), 5.20 (1H, dd, $J=10.5, 1.5$ Hz), 5.22 (1H, dd, $J=18, 1.5$ Hz), 6.18 (1H, dd, $J=18, 10.5$ Hz), 6.70 (1H, d, $J=2$ Hz), 6.84 (1H, d, $J=8$ Hz), 6.97 (1H, d, $J=8$ Hz), 8.42 (1H, d, $J=2$ Hz, NH), 10.14 (1H, br s, OH). Ethyl 7-[(*R*)-3,7-Dimethyl-1,6-octadien-3-yl]- α -(hydroxyimino)-4-[[(*S*)-1-(methoxycarbonyl)-2-methylpropyl]methylamino]indole-5-propanoate (**16a**) Colorless syrup. HRMS Calcd for $\text{C}_{30}\text{H}_{43}\text{N}_3\text{O}_5$ 525.3202. Found: 525.3220. IR (CHCl_3) cm^{-1} 1720, 1630. $^1\text{H NMR}$ (90 MHz, 60°C) δ 0.90 (3H, d, $J=6.5$ Hz), 1.16 (3H each, d, $J=6.5$ Hz), 1.18 (3H, t, $J=7$ Hz), 1.36 (3H, s), 1.39 (3H, s), 1.57 (3H, s), 2.28 (1H, dq, $J=9, 6.5, 6.5$ Hz), 3.05 (3H, s), 3.43 (3H, s), 3.48 (1H, d, $J=9$ Hz), 4.12 (1H, d, $J=14.5$ Hz), 4.17 (2H, q, $J=7$ Hz), 4.37 (1H, d, $J=14.5$ Hz), 4.87-5.11 (1H, m), 5.17 (1H, dd, $J=18, 1.5$ Hz), 5.19 (1H, dd, $J=10, 1.5$ Hz), 6.20 (1H, dd, $J=18, 10$ Hz), 6.47 (1H, dd, $J=3.5, 2$ Hz), 6.82 (1H, s), 6.97 (1H, dd, $J=3.5, 3$ Hz), 8.24-9.25 (1H, m, OH), 8.43 (1H, br s, NH). Similarly **15b** (65%) and **16b** (15%) were prepared from **9b** along with a 9% yield recovery of **9b**. Ethyl 7-[(*S*)-3,7-Dimethyl-1,6-octadien-3-yl]- α -(hydroxyimino)-4-[[(*S*)-1-(methoxycarbonyl)-2-methylpropyl]methylamino]indole-3-propanoate (**15b**) Colorless syrup. HRMS Calcd for $\text{C}_{30}\text{H}_{43}\text{N}_3\text{O}_5$ 525.3202. Found: 525.3196. IR (CHCl_3) cm^{-1} 1723, 1628. $^1\text{H NMR}$ (90 MHz) δ 0.94 (3H, d, $J=6.5$ Hz), 1.10 (3H, d, $J=6.5$ Hz), 1.19 (3H, t, $J=7$ Hz), 1.41 (6H, s), 1.61 (3H, s), 2.89 (3H, s), 3.57 (3H, s), 3.69 (1H, d, $J=9$ Hz), 4.21 (2H, q, $J=7$ Hz), 4.38 (1H, d, $J=16.5$ Hz), 4.56 (1H, d, $J=16.5$ Hz), 4.86-5.19 (1H, m), 5.19 (1H, dd, $J=10, 1$ Hz), 5.22 (1H, dd, $J=18, 1$ Hz), 6.18 (1H, dd, $J=18, 10$ Hz), 6.70 (1H, d, $J=2$ Hz), 6.86 (1H, d, $J=8$ Hz), 6.98 (1H, d, $J=8$ Hz), 8.42 (1H, s, $J=2$ Hz, NH), 10.40 (1H, br s, OH). Ethyl 7-[(*S*)-3,7-Dimethyl-1,6-octadien-3-yl]- α -(hydroxyimino)-4-[[(*S*)-1-(methoxycarbonyl)-2-methylpropyl]methylamino]indole-5-propanoate (**16b**) Colorless syrup. HRMS Calcd for $\text{C}_{30}\text{H}_{43}\text{N}_3\text{O}_5$ 525.3202. Found: 525.3203. IR (CHCl_3) cm^{-1} 1720, 1629. $^1\text{H NMR}$ (90 MHz, 60°C) δ 0.90 (3H, d, $J=6.5$ Hz), 1.18 (3H, d, $J=6.5$ Hz), 1.21 (3H, t, $J=7$ Hz), 1.37 (3H, s), 1.42 (3H, s), 1.60 (3H, s), 3.07 (3H, s), 3.42 (3H, s), 4.12 (1H, d, $J=15$ Hz), 4.39 (1H, d, $J=15$ Hz), 4.20 (2H, q, $J=7$ Hz), *ca* 4.89-5.15 (1H, m), 5.18 (1H, dd, $J=18, 1.5$ Hz), 5.20 (1H, dd, $J=10.5, 1.5$ Hz), 6.20 (1H, dd, $J=18, 10.5$ Hz), 6.48 (1H, d, $J=3, 2$ Hz), 6.83 (1H, s), 6.98 (1H, dd, $J=3, 3$ Hz), 8.42 (1H, br s, NH), 8.78 (1H, br s, OH).

Aluminium-Amalgam Reduction to Form 17a and 17b — Preparation of **17a** is typical. To a solution of **15a** (86 mg, 0.164 mmol) in $\text{THF-H}_2\text{O}$ (9 l) (8 ml) was added Al-Hg prepared from Al (176 mg), and the mixture was stirred at 18°C for 4 h. The whole was passed through a celite bed and the celite was washed with CH_2Cl_2 . Usual work-up of the combined filtrate and purification by PTLC (3% EtOH- CH_2Cl_2) afforded ethyl 7-[(*R*)-3,7-dimethyl-1,6-octadien-3-yl]-4-[[(*S*)-1-(methoxycarbonyl)-2-methylpropyl]methylamino]tryptophanate (**17a**) (77 mg, 92%), colorless syrup. HRMS Calcd for $\text{C}_{30}\text{H}_{45}\text{N}_3\text{O}_4$ 511.3409. Found: 511.3401. IR (CHCl_3) cm^{-1} 1723, 1628. $^1\text{H NMR}$ (90 MHz) of two diastereomers (*ca* 1:1) δ 1.37 (3H, s), 1.40 (3H, s), 1.58 (3H, s), 1.71 (2H, s, NH_2), 2.85 (3H, s), 3.40 (3H, s), 4.07 and 4.15 (2H, q each, $J=7$ Hz), 4.90-5.19 (1H, m), 5.09-5.41 (2H, m), 6.21 (1H, dd, $J=18, 10.5$ Hz), 6.83 (1H, d, $J=8$ Hz), 6.97 (1H, d, $J=8$ Hz), 6.90-7.00 (1H, m), 8.56 (1H, br s, indole NH). Ethyl 7-[(*S*)-3,7-Dimethyl-1,6-octadien-3-yl]-4-[[(*S*)-1-(methoxycarbonyl)-2-methylpropyl]methylamino]tryptophanate (**17b**) Colorless syrup. HRMS Calcd for $\text{C}_{30}\text{H}_{45}\text{N}_3\text{O}_4$ 511.3409. Found: 511.3391. IR (CHCl_3) cm^{-1} 1723, 1628. $^1\text{H NMR}$ (90 MHz) of two diastereomers (*ca* 1:1) δ 0.94, 0.96, 1.10 and 1.12 (6H, d each, $J=6.5$ Hz), 1.21 (3H, t, $J=7$ Hz), 1.40 (3H, s), 1.44 (3H, s), 1.61 (3H, s), 1.75 (2H, s, NH_2), 2.85 (3H, s), 3.45 (3H, s), 4.09 and 4.16 (2H, q each, $J=7$ Hz), 4.87-5.23 (1H, m), 5.23 (1H, dd, $J=18, 1.5$ Hz), 5.23 (1H, dd, $J=10.5, 1.5$ Hz), 6.19 (1H, dd, $J=18, 10.5$ Hz), 6.83 (1H, d, $J=8$ Hz), 6.91-7.01 (1H, m), 6.97 (1H, d, $J=8$ Hz), 8.55 (1H, br s, indole NH).

Amino Alcohols 18a and 18b — Reduction of **17a** is typical. A mixture of **17a** (28 mg, 0.055 mmol) and NaBH_4 (17 mg, 0.447 mmol) in EtOH was refluxed with stirring under Ar atmosphere for 20 h. After cooling, brine was added and the whole was extracted thoroughly with CH_2Cl_2 . Usual work-up and Al_2O_3 PTLC (4%

MeOH-CH₂Cl₂) gave methyl *N*-[3-(2-amino-3-hydroxypropyl)-7-[(*R*)-3,7-dimethyl-1,6-octadien-3-yl]-4-indolyl]-*N*-methyl-L-valinate (**18a**) (13 mg, 51%), colorless film HRMS Calcd for C₂₈H₄₃N₃O₃, 469.3304 Found 469.3303. IR (CHCl₃) cm⁻¹ 1727, 1628. ¹H NMR (90 MHz) of two diastereomers (*ca* 1:1) δ 0.90 (3H, d, J=6.5 Hz), 1.11 and 1.15 (3H, d each, J=6.5 Hz), 1.40 (3H, s), 1.45 (3H, s), 1.63 (3H, s), *ca* 2.64-3.03 (6H, m, changed to 2.82 and 2.84 (3H, s each) with D₂O), 3.40 and 3.44 (3H, s each), 4.90-5.19 (1H, m), 5.24 (1H, dd, J=10.5, 1 Hz), 5.26 (1H, dd, J=18, 1 Hz), 6.22 (1H, dd, J=18, 10.5 Hz), 6.80 (1H, d, J=8 Hz), 6.90-7.01 (1H, m), 6.98 (1H, d, J=8 Hz), 8.56 (1H, br s, indole NH) Methyl *N*-[3-(2-Amino-3-hydroxypropyl)-7-[(*S*)-3,7-dimethyl-1,6-octadien-3-yl]-4-indolyl]-*N*-methyl-L-valinate (**18b**). Colorless film HRMS Calcd for C₂₈H₄₃N₃O₃, 469.3304. Found 469.3287. IR (CHCl₃) cm⁻¹ 1728, 1630. ¹H NMR (90 MHz) of two diastereomers (*ca* 1:1) δ 0.93 (3H, d, J=6.5 Hz), 1.07 and 1.11 (3H, d each, J=6.5 Hz), 1.41 (3H, s), 1.43 (3H, s), 1.62 (3H, s), 2.80 (3H, s), 3.30 (3H, br s, NH₂ and OH), 3.45 and 3.47 (3H, s each), 4.92-5.21 (1H, m), 5.10-5.40 (2H, m), 6.20 (1H, dd, J=18, 10.5 Hz), 6.80 (1H, d, J=8 Hz), 6.92-7.04 (1H, m, changed to 6.97 (1H, s) with D₂O), 6.98 (1H, d, J=8 Hz), 8.56 (1H, br s, indole NH)

Lyngbyatoxin A (Teleocidin A-1) (1) — A solution of **18a** (14 mg, 0.030 mmol) in 10% KOH in MeOH-H₂O (4:1) (1.5 ml, 2.68 mmol) was refluxed with stirring under Ar atmosphere for 22 h. After cooling at 0°C, Et₃N HCl (363 mg, 2.64 mmol) was added and the mixture was stirred at 0°C for 1 min and then at 20°C for 10 min. The reaction mixture was concentrated *in vacuo* at room temperature and the residue was dried over P₂O₅ in reduced pressure for 1 h. It was suspended in DMF (3 ml) and to this was added successively 5% w/v DPPA/DMF (0.20 ml, 0.036 mmol) and 8% v/v Et₃N/DMF (0.10 ml, 0.058 mmol) at 0°C. The mixture was stirred at 0°C for 1 h and at room temperature for 17 h. The reaction was quenched with sat. NaHCO₃-H₂O, the whole was extracted with Et₂O and worked up as usual. PTLC [hexane-EtOAc (2:3)] afforded a crude mixture of **1** and ent-**2** (4.5 mg) as a less polar fraction and a crude mixture of 9-epi-**1** and 9-epi-ent-**2** (4 mg) as a more polar fraction. Both were purified respectively by PTLC (5% MeOH-CH₂Cl₂) to give a mixture of **1** and ent-**2** (3.5 mg) and a mixture of 9-epi-**1** and 9-epi-ent-**2** (3.5 mg, 27%). The former was separated by HPLC [column YMC A-024, SIL, S-5, 10 × 300 mm, mobile phase hexane-CH₂Cl₂-2-propanol (85:10:5), flow rate 4 ml/min] to afford lyngbyatoxin A (teleocidin A-1) (**1**) (3 mg, 23%, retention time 16.8 min) and (+)-teleocidin A-2 (ent-**2**) (0.5 mg, *ca* 4%, retention time 18.5 min). The synthetic **1** Colorless film HRMS Calcd for C₂₇H₃₉N₃O₂, 437.3042. Found 437.3028. CD (c 1.67 × 10⁻⁴, MeOH, 21°C) [θ]₃₃₃ 0, [θ]₃₁₃ +4600, [θ]₂₉₈ 0, [θ]₂₅₇ -17700, [θ]₂₃₇ 0, [θ]₂₃₄ +7200, [θ]₂₂₉ 5, [θ]_{221.5} -9300, [θ]₂₁₅ 0. IR (CHCl₃) cm⁻¹ 1657. ¹H NMR (400 MHz, CDCl₃-D₂O) δ (major conformer) 0.64 (3H, d, J=6.5 Hz), 0.92 (3H, d, J=6.5 Hz), 1.47 (3H, s), 1.51 (3H, s), 1.66 (3H, s), 1.67-2.06 (4H including minor conformer, m), 2.58 (1H, dq, J=10.5, 6.5, 6.5 Hz), 2.90 (3H, s), 3.15 (1H, br d, J=17 Hz), 3.56 (1H, dd, =11, 8.5 Hz), 3.73 (1H, dd, J=11, 3.5 Hz), 4.29-4.36 (1H, m), 4.34 (1H, d, J=10.5 Hz), 5.02-5.11 (1H, m), 5.28 (1H, d, J=11 Hz), 5.30 (1H, d, J=18 Hz), 6.17 (1H, dd, J=18, 11 Hz), 6.47 (1H, d, J=8 Hz), 6.83 (1H, s), 6.97 (1H, d, J=8 Hz), 8.52 (1H, s, indole NH), (minor conformer) 0.93 (3H, d, J=6.5 Hz), 1.25 (3H, d, J=6.5 Hz), 1.45 (3H, s), 1.49 (3H, s), 1.64 (3H, s), 2.38 (1H, dq, J=11, 6.5, 6.5 Hz), 2.73 (3H, s), 2.80 (1H, dd, J=14.5, 1.5 Hz), 3.39 (1H, dd, J=11, 7 Hz), 3.46 (1H, dd, =11, 6.5 Hz), 4.40-4.47 (1H, m), 5.02-5.11 (1H, m), 5.33 (1H, d, J=11 Hz), 5.35 (1H, d, J=18 Hz), 6.22 (1H, dd, J=18, 11 Hz), 6.96 (1H, s), 7.01 (1H, d, J=8 Hz), 7.09 (1H, d, J=8 Hz), 8.76 (1H, s, indole NH). ¹³C NMR (100 MHz, CDCl₃-D₂O) δ 17.5, 19.6, 21.6, 23.1, 24.2, 25.7, 28.5, 33.1, 33.9, 38.6, 43.3, 55.7, 65.0, 71.0, 106.5, 112.4, 114.0, 118.7, 120.0, 120.9, 121.6, 124.6, 131.5, 137.6, 146.4, 148.4, 174.4. (+)-Teleocidin A-2 (ent-**2**) Colorless film HRMS Calcd for C₂₇H₃₉N₃O₂, 437.3042. Found 437.3041. CD (c 1.05 × 10⁻⁴, MeOH, 22°C) [θ]₃₂₉ 0, [θ]₃₁₄ -3200, [θ]₃₀₂ 0, [θ]_{252.5} +14800, [θ]_{249.5} +14300, [θ]₂₂₇ +50500, [θ]_{212.5} 0. ¹H NMR spectrum (270 MHz) was identical with that of teleocidin A-2 (**2**) described below. 9-Epi-**1** and 9-epi-ent-**2** Colorless film. Retention time of HPLC [TSK Silica 60, hexane-CHCl₃-2-propanol (85:10:5), 1 ml/min] 20.8 min. HRMS Calcd for C₂₇H₃₉N₃O₂, 437.3042. Found 437.3041. IR (CHCl₃) cm⁻¹ 1654. ¹H NMR (270 MHz, CDCl₃-D₂O) of major conformer δ 1.47 (3H, s), 1.49 (3H, s), 1.64 (3H, s), 3.09 (3H, br s), 5.02-5.11 (1H, m), 6.17 (1H, dd, J=18, 11 Hz), 6.81 (1H, s), 8.42 (1H, br s, indole NH)

Teleocidin A-2 (2) — The same treatment of **18b** (13 mg, 0.028 mmol) as above afforded teleocidin A-2 (**2**) (2.5 mg, 21%), (+)-teleocidin A-1 (ent-**1**) (0.5 mg, *ca* 4%), and a mixture of 9-epi-**2** and 9-epi-ent-**1** (3 mg, 25%). Synthetic teleocidin A-2 (**2**). Colorless film HRMS Calcd for C₂₇H₃₉N₃O₂, 437.3042. Found 437.3017. CD (c 2.72 × 10⁻⁴, MeOH, 23°C) [θ]₃₃₆ 0, [θ]₃₁₄ +3900, [θ]₃₀₂ 0, [θ]₂₅₆ -17500, [θ]_{250.5} -17000, [θ]₂₂₇ -58000, [θ]₂₁₂ 0. IR (CHCl₃) cm⁻¹ 1658. ¹H NMR (400 MHz, CDCl₃-D₂O) δ (major conformer) 0.60 (3H, d, J=6.5 Hz), 0.91 (3H, d, J=6.5 Hz), 1.38 (3H, s), 1.45 (3H, s), 1.63 (3H, s), 1.76-2.04 (4H including minor conformer, m), 2.58 (1H, dq, J=10.5, 6.5, 6.5 Hz), 2.90 (3H, s), 3.03 (1H, dd, J=17, 3.5 Hz), 3.15 (1H, br d, J=17 Hz), 3.54 (1H,

dd, $J=11.5, 8.5$ Hz), 3.72 (1H, dd, $J=11.5, 3.5$ Hz), 4.30-4.37 (1H, m), 4.34 (1H, d, $J=10.5$ Hz), 5.07 (1H of major and minor conformers, br dd, $J=6.5, 6.5$ Hz), 5.26 (1H, d, $J=11$ Hz), 5.30 (1H, d, $J=18$ Hz), 6.20 (1H of major and minor conformers, dd, $J=18, 11$ Hz), 6.47 (1H, d, $J=8$ Hz), 6.82 (1H, br s), 6.97 (1H d, $J=8$ Hz), 8.53 (1H, br s, indole NH), (minor conformer) 0.94 (3H, d, $J=6.5$ Hz), 1.25 (3H, d, $J=6.5$ Hz), 1.48 (3H, s), 1.65 (3H, s), 2.39 (1H, dq, $J=11, 6.5, 6.5$ Hz), 2.74 (3H, s), 2.80 (1H, dd, $J=15, 1.5$ Hz), 3.38 (1H, dd, $J=11, 7$ Hz), 3.46 (1H, dd, $J=11, 6$ Hz), 4.40-4.46 (1H, m), 5.33 (1H, d, $J=11$ Hz), 5.34 (1H, d, $J=18$ Hz), 7.01 (1H, d, $J=8$ Hz), 7.10 (1H d, $J=8$ Hz), 8.76 (1H, br s, indole NH) ^{13}C NMR (100 MHz, $\text{CDCl}_3\text{-D}_2\text{O}$) δ : 17.3, 19.4, 21.6, 23.1, 24.8, 25.6, 28.5, 33.0, 34.0, 38.0, 43.3, 55.5, 65.0, 71.0, 106.0, 112.1, 114.0, 118.5, 120.3, 120.9, 121.1, 124.6, 131.4, 137.5, 146.5, 149.0, 174.2 (+)-Teleocidin A-1 (ent-1). Colorless film HRMS Calcd for $\text{C}_{27}\text{H}_{39}\text{N}_3\text{O}_2$ 437.3042 Found 437.3018. CD (c 1.37×10^{-4} , MeOH, 21°C) $[\theta]_{333}^0$, $[\theta]_{312}^0 -4600$, $[\theta]_{298}^0$, $[\theta]_{258}^0 +17500$, $[\theta]_{238}^0$, $[\theta]_{233}^0 -9100$, $[\theta]_{228.5}^0$, $[\theta]_{222}^0 +8800$. ^1H NMR spectrum (270 MHz) was identical with that of teleocidin A-1 (1) described above 9-Epi-2 and 9-epi-ent-1 Colorless film HRMS Calcd for $\text{C}_{27}\text{H}_{39}\text{N}_3\text{O}_2$ 437.3042 Found 437.3052 IR (CHCl_3) cm^{-1} 1655 ^1H NMR (270 MHz) δ 0.67 (3H, d, $J=6.5$ Hz), 0.74 (3H, d, $J=6.5$ Hz), 1.40 (3H, s), 1.44 (3H, s), 1.63 (3H, s), 2.89 (1H, br d, $J=15.5$ Hz), 3.09 (3H, s), 3.28 (1H, br d, $J=15.5$ Hz), 5.02-5.12 (1H, m), 5.26 (1H, dd, $J=11, 1.5$ Hz), 5.28 (1H, dd, $J=18, 1.5$ Hz), 6.21 (1H, dd, $J=18, 11$ Hz), 6.73 (1H, d, $J=8$ Hz), 6.80 (1H, s), 6.95 (1H, d, $J=8$ Hz), 7.32 (1H, br s, CONH), 8.44 (1H, br s, indole NH)

(+)-MTPA Esters of Teleocidin A-1 (1), Teleocidin A-2 (2), (+)-Teleocidin A-1 (ent-1) and (+)-Teleocidin A-2 (ent-2) — Preparation of (+)-MTPA ester of teleocidin A-2 (2) is typical. A solution of 2 (3.5 mg, 0.008 mmol) in pyridine (0.2 ml) and CH_2Cl_2 (0.2 ml) was stirred with the acid chloride (24 mg, 0.095 mmol) derived from (*R*)-(+)-MTPA at 0°C for 1 h. Sat. $\text{NaHCO}_3\text{-H}_2\text{O}$ was added, the mixture was extracted with Et_2O and the extract was worked up as usual. Purification by PTLC [hexane-EtOAc (6:1)] afforded a crude product (5 mg), which was further purified by PTLC [hexane- CH_2Cl_2 (2:3)] to give the (+)-MTPA ester of 2 (4.5 mg, 86%), colorless film MS m/z 653 (M^+) IR (CHCl_3) cm^{-1} 1752, 1667 ^1H NMR (90 MHz) of two conformers δ 2.71 and 2.83 (3H, s each), 3.51 (3H, s), 6.79 (1H, br s), 6.96 (1H, d, $J=7.5$ Hz), 7.10 (1H, d, $J=7.5$ Hz), 8.53 and 8.71 (1H, br s each, indole NH). The (+)-MTPA ester of 1. Colorless film MS m/z 653 (M^+) IR (CHCl_3) cm^{-1} 1750, 1670 ^1H NMR (90 MHz) of major and minor conformers δ 2.69 and 2.83 (3H, s each), 3.51 (3H, s), 6.79 (1H, br s), 6.97 (1H, d, $J=7.5$ Hz), 7.09 (1H, d, $J=7.5$ Hz), 8.76 and 8.53 (1H, br s each, indole NH). In the same manner as above, ent-1 (*ca* 0.3 mg) and ent-2 (*ca* 0.5 mg) were converted to their (+)-MTPA esters (*ca* 0.3 mg and *ca* 0.5 mg)

HPLC Analysis of the Four (+)-MTPA Esters — Column YMC A-314 (ODS, 6×300 mm), mobile phase MeOH- H_2O (88:12), flow rate 3 ml/min. Retention time of (+)-MTPA esters of 1, 2, ent-1 and ent-2 39.1 min, 36.4 min, 40.3 min and 37.6 min

Methyl *N*-[5,5-Dimethyl-4-hydroxy-4-(2-pyrrolyl)-6-heptenoyl]-*N*-methyl-L-valinate (20a) and Methyl *N*-[(*E*)-4-Hydroxy-4-(2-pyrrolyl)-5,9,13-trimethyl-5-vinyl-8,12-tetradecadienoyl]-*N*-methyl-L-valinate (20b) — Grignard reaction of 5 with 3-methyl-2-butenyl (prenyl) bromide or (*E, E*)-3,7,11-trimethyl-2,6,10-dodecatrienyl (farnesyl) bromide was carried out in the same manner as in the previous report¹⁶ at -20°C for 20a or 0°C for 20b. 20a Colorless syrup MS m/z 347 ($M^+ - \text{OH}$) IR (CHCl_3) cm^{-1} 1740, 1635 (sh), 1620 ^1H NMR (90 MHz) of two diastereomers δ 0.70-0.92 (3H, m), 0.73 (3H, d, $J=7$ Hz), 0.98 (6H, s), 2.78 and 2.81 (3H, s each), 3.56-3.71 and 4.74 (1H, m and d, $J=10$ Hz), 3.66 (3H, s), 4.62 (1H, s, OH), 5.71-5.90 (1H, m), 6.04-6.22 (1H, m), 6.57-6.73 (1H, m), 8.87 (1H, br s, NH). 20b Colorless syrup MS m/z 482 ($M^+ - \text{H}_2\text{O}$) IR (CHCl_3) cm^{-1} 1738, 1620 ^1H NMR (90 MHz) δ 0.75, 0.80 and 0.96 (6H, d each, $J=7$ Hz), 1.01 (3H, s), 1.52 (3H, s), 1.60 (3H, s), 1.67 (3H, s), 2.78 and 2.81 (3H, s each), 3.58 and 3.66 (3H, s each), 3.77 and 4.87 (1H, d each, $J=10.5$ Hz), 4.12-4.35 (1H, m, OH), 5.70-5.88 (1H, m), 6.05-6.21 (1H, m), 6.54-6.71 (1H, m), 8.73 (1H, br s, NH)

Methyl *N*-[5,5-Dimethyl-4-(2-pyrrolyl)-3,6-heptadienoyl]-*N*-methyl-L-valinate (21a) and Methyl *N*-Methyl-*N*-[(3 ξ , 8*E*)-4-(2-pyrrolyl)-5,9,13-trimethyl-5-vinyl-3,8,12-tetradecatrienoyl]-L-valinate (21b) — In the same manner as in the previous paper,¹⁶ 20a and 20b were dehydrated with a catalytic amount of *p*-TsOH to yield 21a and 21b. 21a Colorless syrup MS m/z 346 (M^+) IR (CHCl_3) cm^{-1} 1740, 1635 ^1H NMR (90 MHz) of major and minor rotamers δ 0.85 and 0.83 (3H, d each, $J=7$ Hz), 1.00 and 0.94 (3H, d each, $J=7$ Hz), 1.18 (6H, s), 1.92-2.05 (1H, m), 2.92 and 2.87 (3H, s each), 3.07 and 3.13 (3H, d each, $J=7.5$ Hz), 3.70 (3H, s), 4.91 and 3.90 (1H, d each, $J=10.5$ Hz), 4.93-5.21 (2H, m), 5.80-6.23 (3H, m), 6.63-6.75 (1H, m), 9.28 (1H, br s, NH). 21b Colorless syrup MS m/z 482 (M^+) IR (CHCl_3) cm^{-1} 1738, 1632 ^1H NMR (90 MHz) δ 0.81, 0.85 and 1.00 (6H, d each, $J=7$ Hz), 1.20 (3H, s), 1.51 (3H, s), 1.58 (3H, s), 1.66 (3H, s), 2.86 and 2.91 (3H, s each),

3 06, 3 07 and 3 12 (2H, d each, $J=7.5$ Hz), 3 69 (3H, s), 5 86 (1H, t, $J=7.5$ Hz), 6 57-6.77 (1H, m), 9 31 (1H, br s, NH)

Methyl *N*-[5,5-Dimethyl-4-(2-pyrrolyl)-3,6-heptadienthioyl]-*N*-methyl-L-valinate (22a) and Methyl *N*-Methyl-*N*-[(3 ξ , 8*E*)-4-(2-pyrrolyl)-5,9,13-trimethyl-5-vinyl-3,8,12-tetradecatrienthioyl]-L-valinate (22b) — Thioamides 22a and 22b were prepared by the reaction of 21a and 21b with Lawesson's reagent as in the previous paper.¹⁶ 22a Colorless syrup MS m/z : 362 (M^+). IR (CHCl₃) cm^{-1} : 1740. ¹H NMR (90 MHz) of two rotamers δ 0.76, 0.91, 0.93 and 1 10 (6H, d each, $J=7$ Hz), 1 16 (6H, s), 2 31 (1H, dq, $J=10.5, 7, 7$ Hz), 3 04 and 3 30 (3H, s each), 3 49 and 3 56 (2H, d each, $J=7.5$ Hz), 4 27 and 5 87 (1H, d each, $J=10.5$ Hz), 5 03 (1H, d, $J=12$ Hz), 5.05 (1H, d, $J=17.5$ Hz), 5.73 and 5.76 (1H, t each, $J=7.5$ Hz), 5 92-6.07 (1H, m), 6.07-6 22 (1H, m), 6 70-6 77 (1H, m), 8 80 (1H, br s, NH) 22b Colorless syrup. MS m/z : 498 (M^+) IR (CHCl₃) cm^{-1} : 1737 ¹H NMR (90 MHz) of two rotamers δ 0 77, 0 90, 0 91 and 1 08 (6H, d each, $J=7$ Hz), 1 21 (3H, s), 1 55 (3H, s), 1 59 (3H, s), 1 67 (3H, s), 3 05 and 3 31 (3H, s each), 3 71 (3H, s), 4.27 and 6.31 (1H, d each, $J=10.5$ Hz), 5 70 and 5 73 (1H, t each, $J=7.5$ Hz), 5 86-6 23 (2H, m), 6 56-6 78 (1H, m), 8 81 (1H, br s, NH)

Indole Cyclization with MeI — In a similar manner as in the preceding paper,¹ 22a and 22b were cyclized to indole derivatives 23a, 23b and 24b with MeI in DMF **Methyl *N*-[7-(1,1-Dimethyl-2-propenyl)-4-indolyl]-*N*-methyl-L-valinate (23a)**: Colorless needles, mp 100-103°C or colorless prisms, mp 106-107°C (MeOH-H₂O) Anal Calcd for C₂₀H₂₄N₂O₂ C, 73 14, H, 8 59, N, 8 53 Found. C, 73.00, H, 8 55, N, 8 38 [α]_D²³ -44 1° (c 0 503, CHCl₃) MS m/z : 328 (M^+) IR (KBr) cm^{-1} : 1720 ¹H NMR (90 MHz) δ 0 94 (3H, d, $J=7$ Hz), 1 07 (3H, d, $J=7$ Hz), 1 50 (6H, s), 2 37 (1H, dq, $J=11, 7, 7$ Hz), 3 04 (3H, s), 3.66 (3H, s), 4.08 (1H, d, $J=11$ Hz), 5 20 (1H, dd, $J=10.5, 1.5$ Hz), 5.29 (1H, dd, $J=18, 1.5$ Hz), 6 25 (1H, dd, $J=18, 10.5$ Hz), 6 60 (1H, d, $J=8$ Hz), 6 71 (1H, dd, $J=3, 2$ Hz), 7 03 (1H, d, $J=8$ Hz), 7 07 (1H, dd, $J=3, 3$ Hz), 8 60 (1H, br s, NH) **7-(1,1-Dimethyl-2-propenyl)-4-(methylthio)indole (25a)** Colorless needles, mp 82.5-83°C (MeOH-H₂O) Anal Calcd for C₁₄H₁₇NS C, 72 68, H, 7 41, N, 6 05 Found. C, 72 46, H, 7 54, N, 6 10 MS m/z : 231 (M^+) IR (KBr) cm^{-1} : 1630 ¹H NMR (90 MHz) δ 1 50 (6H, s), 2 54 (3H, s), 5 18 (1H, dd, $J=10.5, 1.5$ Hz), 5 27 (1H, dd, $J=18, 1.5$ Hz), 6 21 (1H, dd, $J=18, 10.5$ Hz), 6 62 (1H, dd, $J=3, 2$ Hz), 6 96 (1H, d, $J=8$ Hz), 7 05-7 20 (1H, m), 7 11 (1H, d, $J=8$ Hz), 8 63 (1H, br s, NH) **Methyl *N*-Methyl-*N*-[7-(*R*, *E*)-3,7,11-trimethyl-1,6,10-dodecatrien-3-yl]-4-indolyl]-L-valinate (23b)** Colorless syrup HRMS Calcd for C₃₀H₄₄N₂O₂ 464 3403. Found. 464 3384 IR (CHCl₃) cm^{-1} : 1725 ¹H NMR (400 MHz) δ : 0 94 (3H, d, $J=7$ Hz), 1 10 (3H, d, $J=7$ Hz), 1 43 (3H, s), 1 46 (3H, s), 1 59 (3H, s), 1 67 (3H, s), 2 38 (1H, dq, $J=11, 7, 7$ Hz), 2 99 (3H, s), 3 59 (3H, s), 4.07 (1H, d, $J=11$ Hz), 5 00-5 14 (2H, m), 5 26 (1H, dd, $J=11, 1$ Hz), 5 29 (1H, dd, $J=18, 1$ Hz), 6 22 (1H, dd, $H=18, 11$ Hz), 6 57 (1H, d, $J=8$ Hz), 6 68 (1H, dd, $J=3, 2$ Hz), 6 98 (1H, d, $J=8$ Hz), 7 07 (1H, dd, $J=3, 3$ Hz), 8 62 (1H, br s, NH) **Methyl *N*-Methyl-*N*-[7-[(*S*, *E*)-3,7,11-trimethyl-1,6,10-dodecatrien-3-yl]-4-indolyl]-L-valinate (24b)** Colorless syrup HRMS Calcd for C₃₀H₄₄N₂O₂ 464 3403 Found 464 3401 IR (CHCl₃) cm^{-1} : 1725 ¹H NMR (400 MHz) δ 0 93 (3H, d, $J=7$ Hz), 1 04 (3H, d, $J=7$ Hz), 1 44 (3H, s), 1 46 (3H, s), 1 58 (3H, s), 1 67 (3H, s), 2 38 (1H, dq, $J=11, 7, 7$ Hz), 3 02 (3H, s), 3 66 (3H, s), 4 08 (1H, d, $J=11$ Hz), 5 02-5 13 (2H, m), 5 26 (1H, dd, $J=11, 1.5$ Hz), 5 28 (1H, dd, $J=18, 1.5$ Hz), 6 22 (1H, dd, $H=18, 11$ Hz), 6 57 (1H, d, $J=8$ Hz), 6 71 (1H, dd, $J=3, 2$ Hz), 6 97 (1H, d, $J=8$ Hz), 7 06 (1H, dd, $J=3, 3$ Hz), 8 61 (1H, br s, NH) **4-(Methylthio)-7-[(*E*)-3,7,11-trimethyl-1,6,10-dodecatrien-3-yl]indole (25b)** Colorless syrup HRMS Calcd for C₂₄H₃₃NS 367 2334 Found 367 2315 ¹H NMR (400 MHz) δ 1 45 (3H, s), 1 48 (3H, s), 1 59 (3H, s), 1 67 (3H, d, $J=1$ Hz), 2 56 (3H, s), 5 02-5 13 (2H, m), 5 29 (1H, dd, $J=11, 1.5$ Hz), 5 30 (1H, dd, $J=18, 1.5$ Hz), 6 22 (1H, dd, $H=18, 11$ Hz), 6 62 (1H, dd, $J=3, 2$ Hz), 6 98 (1H, d, $J=8$ Hz), 7 09 (1H, d, $J=8$ Hz), 7 14 (1H, dd, $J=3, 3$ Hz), 8 66 (1H, br s, NH)

Reaction of 23a, 23b and 24b with Ethyl 3-Bromo-2-hydroxyiminopropanoate — According to the reported procedure,¹ 26a, 26b and 27b were prepared together with some by-products **Ethyl 7-(1,1-Dimethyl-2-propenyl)- α -(hydroxyimino)-4-[[(*S*)-1-(methoxycarbonyl)-2-methylpropyl]methylamino]indole-3-propanoate (26a)** Colorless syrup MS m/z : 457 (M^+) IR (CHCl₃) cm^{-1} : 1725 ¹H NMR (90 MHz) δ : 0 94 (3H, d, $J=7$ Hz), 1 13 (3H, d, $J=7$ Hz), 1 22 (3H, t, $J=7$ Hz), 1 47 (6H, s), 2 00-2 53 (1H, m), 2 92 (3H, s), 3 60 (3H, s), 3 68 (1H, d, $J=9$ Hz), 4 22 (2H, q, $J=7$ Hz), 4 37 (1H, d, $J=15$ Hz), 4 55 (1H, d, $J=15$ Hz), 5 12 (1H, dd, $J=10.5, 1.5$ Hz), 5 23 (1H, dd, $J=18, 1.5$ Hz), 6 16 (1H, dd, $J=18, 10.5$ Hz), 6 70 (1H, d, $J=2$ Hz), 6 85 (1H, d, $J=8$ Hz), 7 00 (1H, d, $J=8$ Hz), 8 38 (1H, br s, NH), 10 14 (1H, br s, OH) **Ethyl 7-(1,1-Dimethyl-2-propenyl)- α -(hydroxyimino)-4-[[(*S*)-1-(methoxycarbonyl)-2-methylpropyl]methylamino]indole-5-propanoate (28a)** Colorless syrup MS m/z : 457 (M^+) IR (CHCl₃) cm^{-1} : 1722 ¹H NMR (90 MHz) δ 0 92 (3H, d, $J=7$ Hz), 1 23 (3H, d, $J=7$ Hz), 1 25 (3H, t, $J=7$ Hz), 1 46 (6H, s), 2.07-2 56 (1H, m), 3 10 (3H, s), 3 45 (3H, s), 4 12 (1H, d, $J=15$ Hz), 4 24 (2H, q, $J=7$ Hz), 4 43 (1H, d, $J=15$ Hz), 5 16 (1H, dd, $J=10.5, 1.5$ Hz), 5 24 (1H, dd, $J=18, 1.5$ Hz), 6 18

(1H, dd, J=18, 10.5 Hz), 6.46 (1H, dd, J=3, 2 Hz), 6.87 (1H, s), 7.00 (1H, dd, J=3, 3 Hz), 8.46 (1H, br s, NH), 9.65 (1H, br s, OH). **Diethyl 7-(1,1-Dimethyl-2-propenyl)- α,α' -bis(hydroxyimino)-4-[[*(S)*-1-(methoxycarbonyl)-2-methylpropyl]methylamino]indole-3,5-bispropanoate (29a)**. Colorless film MS m/z. 586 (M^+) IR (CHCl₃) cm⁻¹ 1728. ¹H NMR (90 MHz) δ 0.74 (3H, d, J=7 Hz), 0.97 (3H, d, J=7 Hz), 1.24 (3H, t, J=7 Hz), 1.25 (3H, t, J=7 Hz), 1.42 (6H, s), 1.65-2.30 (1H, m), 2.97 and 3.01 (3H, s each), 3.54 and 3.77 (3H, s each), 5.13 (1H, d, J=10.5 Hz), 5.22 (1H, d, J=18 Hz), 6.13 (1H, dd, J=18, 10.5 Hz), 6.53-6.77 (1H, m), 6.63 and 6.77 (1H, s each), 8.24 (1H, br s, NH), 9.24 (2H, br s, OH). **Ethyl 9,9a-Dihydro-8-(1,1-dimethyl-2-propenyl)-3-(ethoxycarbonyl)- α -hydroxyimino-5-[[*(S)*-1-(methoxycarbonyl)-2-methylpropyl]methylamino]-1,2-oxazino[6,5-b]indole-4a(4H)-propanoate (30a)**. Colorless film MS m/z 586 (M^+). IR (CHCl₃) cm⁻¹ 1723. ¹H NMR (90 MHz) δ 0.93 (3H, d, J=7 Hz), 2.68 (3H, s), 3.60 and 3.63 (3H, s each), 5.97 (1H, dd, J=18, 10.5 Hz), 6.51 and 6.54 (1H, d each, J=9 Hz), 6.98 (1H, d, J=9 Hz), 8.71 (1H, br s, OH). **Ethyl α -(Hydroxyimino)-4-[[*(S)*-1-(methoxycarbonyl)-2-methylpropyl]methylamino]-7-[[*(R, E)*]-3,7,11-trimethyl-1,6,10-dodecatrien-3-yl]indole-3-propanoate (26b)**. Colorless syrup. MS m/z 593 (M^+). IR (CHCl₃) cm⁻¹ 1730. ¹H NMR (90 MHz) δ 0.92 (3H, d, J=7 Hz), 1.13 (3H, d, J=7 Hz), 1.22 (3H, t, J=7 Hz), 1.45 (6H, s), 1.59 (3H, s), 1.68 (3H, s), 2.94 (3H, s), 3.53 (3H, s), 3.68 (1H, d, J=9 Hz), 4.23 (2H, q, J=7 Hz), 4.58 (1H, d, J=16 Hz), 4.89-5.23 (2H, m), 5.21 (1H, d, J=10.5 Hz), 5.23 (1H, d, J=18 Hz), 6.19 (1H, dd, J=18, 10.5 Hz), 6.71 (1H, d, J=2 Hz), 6.85 (1H, d, J=8 Hz), 6.98 (1H, d, J=8 Hz), 8.41 (1H, br s, NH), 10.09 (1H, br s, OH). **Ethyl α -(Hydroxyimino)-4-[[*(S)*-1-(methoxycarbonyl)-2-methylpropyl]methylamino]-7-[[*(R, E)*]-3,7,11-trimethyl-1,6,10-dodecatrien-3-yl]indole-5-propanoate (28b; R²=vinyl, R³=Me)**. Colorless syrup MS m/z 593 (M^+) IR (CHCl₃) cm⁻¹ 1720. ¹H NMR (90 MHz) δ 0.91 (3H, d, J=7 Hz), 1.22 (3H, d, J=7 Hz), 1.22 (3H, t, J=7 Hz), 1.38 (3H, s), 1.42 (3H, s), 1.57 (3H, s), 1.65 (3H, s), 3.08 (3H, s), 3.23-3.56 (1H, m), 3.43 (3H, s), 4.17 (2H, q, J=7 Hz), 4.39 (1H, d, J=15 Hz), 4.86-5.21 (2H, m), 5.19 (1H, dd, J=18, 1.5 Hz), 5.20 (1H, dd, J=10.5, 1.5 Hz), 6.19 (1H, dd, J=18, 10.5 Hz), 6.43 (1H, dd, J=3, 2 Hz), 6.80 (1H, s), 6.96 (1H, dd, J=3, 3 Hz), 8.46 (1H, br s, NH), 9.56 (1H, br s, OH). **Diethyl α,α' -Bis(hydroxyimino)-4-[[*(S)*-1-(methoxycarbonyl)-2-methylpropyl]methylamino]-7-[[*(R, E)*]-3,7,11-trimethyl-1,6,10-dodecatrien-3-yl]indole-3,5-bispropanoate (29b; R²=vinyl, R³=Me)**. Colorless film MS m/z 722 (M^+) IR (CHCl₃) cm⁻¹ 1722. ¹H NMR (90 MHz) δ : 0.71 (3H, d, J=7 Hz), 0.96 (3H, d, J=7 Hz), 1.26 (6H, t, J=7 Hz), 1.34 (3H, s), 1.40 (3H, s), 1.60 (3H, s), 1.69 (3H, s), 2.98 (3H, s), 3.78 (3H, s), 4.22 (2H, q, J=7 Hz), 4.24 (2H, q, J=7 Hz), 5.19 (1H, d, J=10.5 Hz), 5.21 (1H, d, J=18 Hz), 6.16 (1H, dd, J=18, 10.5 Hz), 6.51-6.76 (1H, m), 6.61 (1H, s), 8.28 (1H, br s, NH), 9.50 (2H, br s, OH). **Ethyl 9,9a-Dihydro-3-(ethoxycarbonyl)- α -hydroxyimino-5-[[*(S)*-1-(methoxycarbonyl)-2-methylpropyl]methylamino]-8-[[*(R, E)*]-3,7,11-trimethyl-1,6,10-dodecatrien-3-yl]-1,2-oxazino[6,5-b]indole-4a(4H)-propanoate (30b; R²=vinyl, R³=Me)**. Colorless film MS m/z 722 (M^+) IR (CHCl₃) cm⁻¹ 1725. ¹H NMR (90 MHz) δ 0.93 (6H, d, J=7 Hz), 1.33 (3H, s), 1.51 (3H, s), 1.61 (3H, s), 1.67 (3H, s), 2.69 and 2.82 (3H, s each), 3.54 and 3.57 (3H, s each), 4.25 (4H, q, J=7 Hz), 5.97 (1H, dd, J=18, 10.5 Hz), 6.49 and 6.52 (1H, d each, J=8 Hz), 6.93 (1H, d, J=8 Hz), 8.78 (1H, br s, OH). **Ethyl α -(Hydroxyimino)-4-[[*(S)*-1-(methoxycarbonyl)-2-methylpropyl]methylamino]-7-[[*(S, E)*]-3,7,11-trimethyl-1,6,10-dodecatrien-3-yl]indole-3-propanoate (27b)**. Colorless syrup MS m/z 593 (M^+) IR (CHCl₃) cm⁻¹ 1725. ¹H NMR (90 MHz) δ 0.93 (3H, d, J=7 Hz), 1.11 (3H, d, J=7 Hz), 1.20 (3H, t, J=7 Hz), 1.44 (6H, s), 1.58 (3H, s), 1.65 (3H, s), 2.91 (3H, s), 3.57 (3H, s), 3.66 (1H, d, J=8 Hz), 4.20 (2H, q, J=7 Hz), 4.53 (1H, d, J=15 Hz), 4.86-5.20 (2H, m), 5.20 (1H, d, J=18 Hz), 5.20 (1H, d, J=10.5 Hz), 6.16 (1H, dd, J=18, 10.5 Hz), 6.62 (1H, d, J=2 Hz), 6.83 (1H, d, J=8 Hz), 6.95 (1H, d, J=8 Hz), 8.38 (1H, br s, NH), 10.06 (1H, br s, OH). **Ethyl α -(Hydroxyimino)-4-[[*(S)*-1-(methoxycarbonyl)-2-methylpropyl]methylamino]-7-[[*(S, E)*]-3,7,11-trimethyl-1,6,10-dodecatrien-3-yl]indole-5-propanoate (28b; R²=Me, R³=vinyl)**. Colorless syrup MS m/z 593 (M^+) IR (CHCl₃) cm⁻¹ 1720. ¹H NMR (90 MHz) δ 0.89 (3H, d, J=7 Hz), 1.23 (3H, d, J=7 Hz), 1.24 (3H, t, J=7 Hz), 1.37 (3H, s), 1.42 (3H, s), 1.58 (3H, s), 1.65 (3H, s), 3.08 (3H, s), 3.24-3.56 (1H, m), 3.40 (3H, s), 4.08 (1H, d, J=15 Hz), 4.18 (2H, q, J=7 Hz), 4.42 (1H, d, J=15 Hz), 4.87-5.21 (2H, m), 5.19 (1H, d, 10.5 Hz), 5.21 (1H, d, J=18 Hz), 6.17 (1H, dd, J=18, 10.5 Hz), 6.45 (1H, dd, J=3, 2 Hz), 6.80 (1H, s), 6.97 (1H, dd, J=3, 3 Hz), 8.45 (1H, br s, NH), 9.57 (1H, br s, OH). **Diethyl α,α' -Bis(hydroxyimino)-4-[[*(S)*-1-(methoxycarbonyl)-2-methylpropyl]methylamino]-7-[[*(S, E)*]-3,7,11-trimethyl-1,6,10-dodecatrien-3-yl]indole-3,5-bispropanoate (29b; R²=Me, R³=vinyl)**. Colorless film MS m/z 722 (M^+) IR (CHCl₃) cm⁻¹ 1725. ¹H NMR (90 MHz) δ 0.74 (3H, d, J=7 Hz), 0.96 (3H, d, J=7 Hz), 1.21 (3H, t, J=7 Hz), 1.25 (3H, t, J=7 Hz), 1.39 (6H, s), 1.56 (3H, s), 1.65 (3H, s), 2.94 and 3.00 (3H, s each), 3.52 and 3.76 (3H, s each), 5.18 (1H, d, J=10.5 Hz), 5.20 (1H, d, J=18 Hz), 6.15 (1H, dd, J=18, 10.5 Hz), 6.50-6.76 (1H, m), 6.57 (1H, s), 8.26 (1H, br s, NH), 9.15 (2H, br s, OH). **Ethyl 9,9a-Dihydro-3-(ethoxycarbonyl)- α -**

hydroxyimino-5-[[[(S)-1-(methoxycarbonyl)-2-methylpropyl]methylamino]-8-[(S, E)-3,7,11-trimethyl-1,6,10-dodecatrien-3-yl]-1,2-oxazino[6,5-b]indole-4a(4H)-propanoate (30b; R²=Me R³=vinyl): Colorless film MS m/z 722 (M⁺) IR (CHCl₃) cm⁻¹ 1722 ¹H NMR (90 MHz) δ: 0.94 (3H, d, J=7 Hz), 1.32, 1.61 and 1.70 (12H, s each), 2.68 and 2.78 (3H, s each), 3.58 and 3.63 (3H, s each), 4.27 (4H, q, J=7 Hz), 5.91 and 5.96 (1H, dd each, J=18, 10.5 Hz), 6.48 and 6.51 (1H, d each, J=8 Hz), 6.94 (1H, d, J=8 Hz), 8.61 and 9.03 (1H, br s each, OH)

Aluminium-Amalgam Reduction to Form Aminodiester 31a, 31b and 32b — Reduction of 26a, 26b and 27b were carried out as above with 20-30 equivalents of Al-Hg at 45°C for 1 h **Ethyl 7-[(1,1-Dimethyl-2-propenyl)-4-[[[(S)-1-(methoxycarbonyl)-2-methylpropyl]methylamino]tryptophanate (31a):** Colorless syrup MS m/z 443 (M⁺) IR (CHCl₃) cm⁻¹ 1728 ¹H NMR (90 MHz) of two diastereomers (*ca.* 1:1) δ 0.94 (3H, d, J=7 Hz), 1.12 (3H, d, J=7 Hz), 1.22 (3H, t, J=7 Hz), 1.48 (6H, s), 1.66 (2H, s, NH₂), 2.04-2.55 (1H, m), 2.85 (3H, s), 3.48 (3H, s), 4.08 and 4.15 (2H, q each, J=7 Hz), 5.16 (1H, d, J=10.5 Hz), 5.23 (1H, d, J=18 Hz), 6.17 (1H, dd, J=18, 10.5 Hz), 6.81 (1H, d, J=8 Hz), 6.90-7.00 (1H, m), 6.98 (1H, d, J=8 Hz), 8.45 (1H, br s, indole NH) **Ethyl 4-[[[(S)-1-(Methoxycarbonyl)-2-methylpropyl]methylamino]-7-[(R, E)-3,7,11-trimethyl-1,6,10-dodecatrien-3-yl]tryptophanate (31b):** Colorless syrup MS m/z 579 (M⁺) IR (CHCl₃) cm⁻¹ 1725 ¹H NMR (90 MHz) of two diastereomers δ 0.93 (3H, d, J=7 Hz), 1.12 and 1.23 (3H, t each, J=7 Hz), 1.17 (3H, d, J=7 Hz), 1.42 (3H, s), 1.48 (3H, s), 1.60 (3H, s), 1.68 (5H, s, changed to 3H, s with D₂O), 2.89 (3H, s), 3.42 (3H, s), 4.06 and 4.15 (2H, q each, J=7 Hz), 4.86-5.23 (2H, m), 5.22 (1H, d, J=10.5 Hz), 5.23 (1H, d, J=18 Hz), 6.20 (1H, dd, J=18, 10.5 Hz), 6.81 (1H, d, J=8 Hz), 6.81-7.06 (1H, m), 6.95 (1H, d, J=8 Hz), 8.53 (1H, br s, indole NH) **Ethyl 4-[[[(S)-1-(Methoxycarbonyl)-2-methylpropyl]methylamino]-7-[(S, E)-3,7,11-trimethyl-1,6,10-dodecatrien-3-yl]tryptophanate (32b):** Colorless syrup MS m/z 579 (M⁺) IR (CHCl₃) cm⁻¹ 1725 ¹H NMR (90 MHz) of two diastereomers δ 0.95 (3H, d, J=7 Hz), 1.11 (3H, d, J=7 Hz), 1.13 and 1.21 (3H, t each, J=7 Hz), 1.43 (6H, s), 1.56 (3H, s), 1.64 (3H, s), 1.78 (2H, br s, NH₂), 2.85 (3H, s), 3.45 (3H, s), 4.08 and 4.15 (2H, q each, J=7 Hz), 4.85-5.24 (2H, m), 5.22 (1H, d, J=18 Hz), 5.22 (1H, d, J=10.5 Hz), 6.18 (1H, dd, J=18, 10.5 Hz), 6.82 (1H, d, J=8 Hz), 6.82-7.05 (1H, m), 6.96 (1H, d, J=8 Hz), 8.52 (1H, br s, indole NH)

Preparation of Trifluoroacetamides 33 – 36 — A procedure for 33a and 34a is typical To a solution of 31a (126 mg, 0.284 mmol) in THF (3.6 ml) and EtOH (4.8 ml) were added LiCl (241 mg, 5.67 mmol) and NaBH₄ (215 mg, 5.66 mmol) at room temperature under Ar atmosphere The mixture was stirred at that temperature for 21.5 h Water was added at 0°C, it was saturated with NaCl, and the whole was extracted with CH₂Cl₂ Usual work-up afforded a residue (114.5 mg), and to its solution in CH₂Cl₂ (12 ml) was added successively at -78°C Et₃N (1.20 ml, 8.63 mmol) and trifluoroacetic anhydride (0.40 ml, 2.83 mmol) After this solution was stirred at -78°C for 15 min, sat NaHCO₃-H₂O was added and the mixture was extracted with CH₂Cl₂ Usual work-up and separation by PTLC (1.5% MeOH-CH₂Cl₂) gave crude 33a (61.5 mg) and 34a (63 mg) Both were purified separately by PTLC [hexane-DME (5:2)] to afford 33a (57 mg, 40%) and 34a (53 mg, 37.5%) **Methyl N-[7-(1,1-Dimethyl-2-propenyl)-3-[(S)-3-hydroxy-2-(trifluoroacetamido)propyl]-4-indolyl]-N-methyl-L-valinate (33a)** Colorless film MS m/z 497 (M⁺) IR (CHCl₃) cm⁻¹ 1720 ¹H NMR (90 MHz) δ 0.95 (3H, d, J=7 Hz), 1.15 (3H, d, J=7 Hz), 1.49 (6H, s), 2.06-2.70 (1H, m), 2.88 (3H, s), 3.19 (1H, dd, J=15, 7 Hz), 3.44 (3H, s), 3.90-4.36 (1H, m), 5.18 (1H, dd, J=10.5, 1 Hz), 5.24 (1H, dd, 18, 1 Hz), 6.16 (1H, dd, J=18, 10.5 Hz), 6.80 (1H, d, J=8 Hz), 6.98-7.12 (1H, m), 7.00 (1H, d, J=8 Hz), 8.62 (1H, br s, indole NH) **Methyl N-[7-(1,1-Dimethyl-2-propenyl)-3-[(R)-3-hydroxy-2-(trifluoroacetamido)propyl]-4-indolyl]-N-methyl-L-valinate (34a)** Colorless film MS m/z 497 (M⁺) IR (CHCl₃) cm⁻¹ 1720 ¹H NMR (90 MHz) δ 0.96 (3H, d, J=7 Hz), 1.08 (3H, d, J=7 Hz), 1.50 (6H, s), 2.04-2.58 (1H, m), 2.85 (3H, s), 3.17 (1H, dd, J=15, 7 Hz), 3.55 (3H, s), 4.06-4.51 (1H, m), 5.16 (1H, dd, J=10.5, 1 Hz), 5.23 (1H, dd, J=18, 1 Hz), 6.17 (1H, dd, J=18, 10.5 Hz), 6.83 (1H, d, J=7.5 Hz), 6.92-7.03 (1H, m), 7.01 (1H, d, J=7.5 Hz), 7.34 (1H, br d, J=7.5 Hz, NHCOCF₃), 8.55 (1H, br s, indole NH) **Methyl N-[3-[(S)-3-Hydroxy-2-(trifluoroacetamido)propyl]-7-[(R)-3,7,11-trimethyl-1,6,10-dodecatrien-3-yl]-4-indolyl]-N-methyl-L-valinate (33b)** Colorless film MS m/z 633 (M⁺) IR (CHCl₃) cm⁻¹ 1720 ¹H NMR (90 MHz) δ 0.92 (3H, d, J=7 Hz), 1.17 (3H, d, J=7 Hz), 1.38 (3H, s), 1.46 (3H, s), 1.56 (3H, s), 1.65 (3H, s), 2.89 (3H, s), 3.34 (3H, s), 3.85-4.25 (1H, m), 4.25-4.74 (1H, m, OH), 4.80-5.22 (2H, m), 5.22 (1H, d, J=18 Hz), 5.24 (1H, d, J=10.5 Hz), 6.18 (1H, dd, J=18, 10.5 Hz), 6.79 (1H, d, J=8 Hz), 6.96 (1H, d, J=8 Hz), 8.62 (1H, br s, indole NH) **Methyl N-[3-[(R)-3-Hydroxy-2-(trifluoroacetamido)propyl]-7-[(R)-3,7,11-trimethyl-1,6,10-dodecatrien-3-yl]-4-indolyl]-N-methyl-L-valinate (34b)** Colorless syrup MS m/z 633 (M⁺) IR (CHCl₃) cm⁻¹ 1727 ¹H NMR (90 MHz) δ 0.93 (3H, d, J=7 Hz), 1.08 (3H, d, J=7 Hz), 1.38 (3H, s), 1.45 (3H, s), 1.57 (3H, s), 1.66 (3H, s), 2.86 (3H, s), 3.45 (3H, s), 4.00-4.45 (1H, m), 4.87-5.24

(2H, m), 5 20 (1H, d, J=18 Hz), 5 22 (1H, d, J=10.5 Hz), 6 18 (1H, dd, J=18, 10 5 Hz), 6.81 (1H, d, J=8 Hz), 6 90-7 00 (1H, m), 6 96 (1H, d, J=8 Hz), 8 56 (1H, br s, indole NH). **Methyl N-[3-[(S)-3-Hydroxy-2-(trifluoroacetamido)propyl]-7-[(S)-3,7,11-trimethyl-1,6,10-dodecatrien-3-yl]-4-indolyl]-N-methyl-L-valinate (35b)**: Colorless syrup MS *m/z* 633 (*M*⁺). IR (CHCl₃) cm⁻¹ 1720 ¹H NMR (90 MHz) δ 0 95 (3H, d, J=7 Hz), 1 15 (3H, d, J=7 Hz), 1 36 (3H, s), 1 44 (3H, s), 1.58 (3H, s), 1.66 (3H, s), 2.88 (3H, s), 3 38 (3H, s), 3 89-4 77 (1H, m, OH), 4.85-5 25 (2H, m), 5 22 (1H, dd, J=18, 1.5 Hz), 5.25 (1H, dd, J=10 5, 1 5 Hz), 6 19 (1H, dd, J=18, 10 5 Hz), 6 79 (1H, d, J=8 Hz), 6 97 (1H, d, J=8 Hz), 8 60 (1H, br s, indole NH). **Methyl N-[3-[(R)-3-Hydroxy-2-(trifluoroacetamido)propyl]-7-[(S)-3,7,11-trimethyl-1,6,10-dodecatrien-3-yl]-4-indolyl]-N-methyl-L-valinate (36b)**: Colorless syrup MS *m/z* 633 (*M*⁺). IR (CHCl₃) cm⁻¹ 1720 ¹H NMR (90 MHz) δ 0 98 (3H, d, J=7 Hz), 1 06 (3H, d, J=7 Hz), 1 39 (3H, s), 1 45 (3H, s), 1 59 (3H, s), 1 67 (3H, s), 2 87 (3H, s), 3 13 (1H, dd, J=14, 7 Hz), 3 47 (3H, s), 3 98-4 50 (1H, m), 4 86-5 24 (2H, m), 5 22 (1H, dd, J=18, 1 5 Hz), 5 24 (1H, dd, J=10, 1 5 Hz), 6 18 (1H, dd, J=18, 10 Hz), 6 82 (1H, d, J=8 Hz), 6 90-7 01 (1H, m), 6 98 (1H, d, J=8 Hz), 8 56 (1H, br s, indole NH).

Pendolmycin (37a=6) and 12-Epipendolmycin (38a) — A solution of 33a (56 mg, 0 113 mmol) and KOH (180 mg, 3 21 mmol) in MeOH (1 3 ml) and H₂O (0 5 ml) was refluxed with stirring under Ar atmosphere for 22 h. After cooling in an ice bath, Et₃N HCl (426.5 mg, 3 10 mmol) was added and the mixture was stirred at 0°C for 1 min and then at 20°C for 10 min. The mixture was concentrated *in vacuo* and the residue was dried thoroughly over P₂O₅ *in vacuo* for 10 h. The resulting solid was suspended in DMF (3 ml) and to this was added successively 5% w/v DEPC/DMF (0 44 ml, 0 135 mmol) and 5% v/v Et₃N/DMF (0 63 ml, 0 226 mmol). The whole was stirred at 20°C for 45 5 h under Ar atmosphere. The reaction was quenched with sat NaHCO₃-H₂O at 0°C and the mixture was extracted with Et₂O. Usual work-up and separation by PTLC [hexane-EtOAc (1 2)] furnished crude **6** (=37a) (23 5 mg) and **38a** (7 mg) in the order of increasing polarity. Both were separately purified by PTLC (4% MeOH-CH₂Cl₂) to give pendolmycin (**6**) (22 5 mg, 54%) as colorless amorphous powder, whose spectral data have been already reported¹⁶ and **38a** (3 8 mg, 9%), colorless needles, mp 198-200°C (MeOH-H₂O). HRMS Calcd for C₂₂H₃₁N₃O₂ 369 2416 Found 369 2407 IR (KBr) cm⁻¹ 1640 ¹H NMR (400 MHz, CDCl₃-D₂O) δ (major conformer) 0 73 (3H, d, J=7 Hz), 0 75 (3H, d, J=7 Hz), 1 47 (3H, s), 1 51 (3H, s), 2 63 (1H, dq, J=10 5, 7, 7 Hz), 2 91 (1H, dd, J=15, 2 5 Hz), 3 10 (3H, s), 3 28 (1H, dd, J=15, 2 5 Hz), 3 77-3 98 (3H containing minor conformer, m), 3 90 (1H, d, J=10 5 Hz), 5 21 (1H, dd, J=10 5, 1 Hz), 5 31 (1H, dd, J=18, 1 Hz), 6 18 (1H, dd, J=18, 10 5 Hz), 6 75 (1H, d, J=8 Hz), 6 83 (1H, d, J=2 Hz), 6 98 (1H, d, J=8 Hz), 8 40 (1H, br s, indole NH), (minor conformer) 0 94 (3H, d, J=7 Hz), 1 26 (3H, d, J=7 Hz), 1 53 (6H, s), 2 37 (1H, dq, J=11, 7, 7 Hz), 2 73 (3H, s), 2 78 (1H, d, J=14 Hz), 3 03 (1H, d, J=11 Hz), 3 05 (1H, d, J=14 Hz), 5 27 (1H, d, J=10 5 Hz), 5 38 (1H, d, J=18 Hz), 6 22 (1H, dd, J=18, 10 5 Hz), 7 01 (1H, d, J=8 Hz), 7 02 (1H, d, J=2 Hz), 7 13 (1H, d, J=8 Hz), 8 68 (1H, br s, indole NH).

Preparation of 7-[(R, E)-3,7,11-Trimethyl-1,6,10-dodecatrien-3-yl]-(-)-indolactam V(37b=7a) and 38b — Similarly 33b (48 mg, 0 076 mmol) was converted to **7a** (=37b) (21 5 mg, 56%) and **38b** (4 5 mg, 12%) **7a=37b** Colorless film HRMS Calcd for C₃₂H₄₇N₃O₂ 505 3668 Found 505 3661 [α]_D²⁴ -147 2° (c 0 67, CHCl₃) IR (CHCl₃) cm⁻¹ 1655 ¹H NMR (400 MHz, CDCl₃-D₂O) δ (major conformer) 0 65 (3H, d, J=7 Hz), 0 92 (3H, d, J=7 Hz), 1 47 (3H, s), 1 50 (3H, s), 1 59 (3H, s), 1 68 (3H, s), 2 58 (1H, dq, J=10, 7, 7 Hz), 2 90 (3H, s), 3 05 (1H, dd, J=18, 3 Hz), 3 15 (1H, d, J=18 Hz), 3 56 (1H, dd, J=12, 8 Hz), 3 73 (1H, dd, J=12, 4 Hz), 4 27-4 39 (1H, m), 4 34 (1H, d, J=10 Hz), 5 01-5 15 (2H containing minor conformer, m), 5 28 (1H, d, J=11 Hz), 5 30 (1H, d, J=18 Hz), 6 17 (1H, dd, J=18, 11 Hz), 6 47 (1H, d, J=8 Hz), 6 83 (1H, s), 6 97 (1H, d, J=8 Hz), 8 51 (1H, br s, indole NH), (minor conformer) 0 93 (3H, d, J=7 Hz), 1 25 (3H, d, J=7 Hz), 1 44 (3H, s), 1 58 (3H, s), 2 38 (1H, dq, J=11, 7, 7 Hz), 2 73 (3H, s), 2 81 (1H, dd, J=15, 2 Hz), 3 00 (1H, d, J=11 Hz), 3 06 (1H, d, J=15 Hz), 3 38 (1H, dd, J=11, 7 Hz), 3 46 (1H, dd, J=11, 6 Hz), 4 39-4 48 (1H, m), 5 33 (1H, d, J=11 Hz), 5 35 (1H, d, J=18 Hz), 6 22 (1H, dd, J=18, 11 Hz), 7 01 (1H, d, J=8 Hz), 7 09 (1H, d, J=8 Hz), 8 77 (1H, br s, indole NH). **7-[(R, E)-3,7,11-Trimethyl-1,6,10-dodecatrien-3-yl]-12epi-(-)-indolactam V(38b)** Colorless prisms, mp 148-149°C (MeOH-H₂O). HRMS Calcd for C₃₂H₄₇N₃O₂ 505 3668 Found 505 3661 IR (CHCl₃) cm⁻¹ 1658 ¹H NMR (400 MHz, CDCl₃-D₂O) δ (major conformer) 0 67 (3H, d, J=7 Hz), 0 74 (3H, d, J=7 Hz), 1 40 (3H, s), 1 45 (3H, s), 1 58 (3H, s), 1 67 (3H, s), 2 61 (1H, dq, J=10, 7, 7 Hz), 2,89 (1H, d, J=16 Hz), 3 10 (3H, s), 3 28 (1H, d, J=16 Hz), 3 91 (1H, d, J=10 Hz), 5 01-5 15 (2H containing minor conformer, m), 5 27 (1H, dd, J=11, 1 Hz), 5 30 (1H, dd, J=18, 1 Hz), 6 21 (1H, dd, J=18, 11 Hz), 6 73 (1H, d, J=8 Hz), 6 81 (1H, d, J=2 Hz), 6 95 (1H, d, J=8 Hz), 8 46 (1H, br s, indole NH), (minor conformer) 0 95 (3H, d, J=7 Hz), 2 37 (1H, dq, J=11, 7, 7 Hz), 2 72 (3H, s), 2 78 (1H, d, J=14 Hz), 3 03 (1H, d, J=11 Hz), 5 34 (1H, d, J=11 Hz), 5 36 (1H, d, J=18 Hz), 7 00

(1H, d, J=8 Hz), 7.01 (1H, d, J=3 Hz), 7.10 (1H, d, J=8 Hz), 8.73 (1H, br s, indole NH).

Preparation of 7-[(S, E)-3,7,11-Trimethyl-1,6,10-dodecatrien-3-yl]-(-)-indolactam V(39b=7b) and 40b — Similarly **35b** (47 mg, 0.074 mmol) was converted to **7b** (=39b) (17.5 mg, 47%) and **40b** (3.5 mg, 9%) **7b=39b** Colorless film. HRMS Calcd for C₃₂H₄₇N₃O₂ 505.3668. Found: 505.3668. $[\alpha]_D^{24}$ -206.4° (c 0.49, CHCl₃) IR (CHCl₃) cm⁻¹ 1655 ¹H NMR (400 MHz, CDCl₃-D₂O) δ (major conformer) 0.60 (3H, d, J=7 Hz), 0.91 (3H, d, J=7 Hz), 1.39 (3H, s), 1.45 (3H, s), 1.58 (3H, s), 1.67 (3H, s), 2.57 (1H, dq, J=10, 7, 7 Hz), 2.90 (3H, s), 3.05 (1H, dd, J=18, 3 Hz), 3.14 (1H, d, J=18 Hz), 3.56 (1H, dd, J=12, 8 Hz), 3.73 (1H, dd, J=12, 3 Hz), 4.28-4.40 (1H, m), 4.35 (1H, d, J=10 Hz), 5.01-5.15 (2H containing minor conformer, m), 5.26 (1H, dd, J=11, 1 Hz), 5.30 (1H, dd, J=18, 1 Hz), 6.21 (1H, dd, J=18, 11 Hz), 6.47 (1H, d, J=8 Hz), 6.83 (1H, s), 6.97 (1H, d, J=8 Hz), 8.53 (1H, br s, indole NH), (minor conformer) 0.94 (3H, d, J=7 Hz), 1.25 (3H, d, J=7 Hz), 1.49 (3H, s), 1.59 (3H, s), 2.38 (1H, dq, J=11, 7, 7 Hz), 2.73 (3H, s), 2.80 (1H, dd, J=15, 1.5 Hz), 2.99 (1H, d, J=11 Hz), 3.37 (1H, dd, J=11, 7 Hz), 3.45 (1H, dd, J=11, 6 Hz), 4.40-4.48 (1H, m), 7.01 (1H, d, J=8 Hz), 7.10 (1H, d, J=8 Hz), 8.76 (1H, br s, indole NH) **7-[(S, E)-3,7,11-Trimethyl-1,6,10-dodecatrien-3-yl]-12epi-(-)-indolactam V(40b)** Colorless film HRMS Calcd for C₃₂H₄₇N₃O₂ 505.3668. Found: 505.3696 IR (CHCl₃) cm⁻¹ 1655 ¹H NMR (400 MHz, CDCl₃-D₂O) δ (major conformer) 0.72 (3H, d, J=7 Hz), 0.75 (3H, d, J=7 Hz), 1.47 (3H, s), 1.49 (3H, s), 1.59 (3H, s), 1.68 (3H, s), 2.63 (1H, dq, J=10.5, 7, 7 Hz), 2.91 (1H, d, J=15 Hz), 3.10 (3H, s), 3.27 (1H, d, J=15 Hz), 3.90 (1H, d, J=10.5 Hz), 5.02-5.14 (2H containing minor conformer, m), 5.28 (1H, d, J=11 Hz), 5.30 (1H, d, J=18 Hz), 6.17 (1H, dd, J=18, 11 Hz), 6.74 (1H, d, J=8 Hz), 6.81 (1H, s), 6.96 (1H, d, J=8 Hz), 8.43 (1H, br s, indole NH), (minor conformer) 0.95 (3H, d, J=7 Hz), 1.26 (3H, d, J=7 Hz), 1.45 (3H, s), 1.50 (3H, s), 2.37 (1H, dq, J=11, 7, 7 Hz), 2.72 (3H, s), 2.78 (1H, d, J=14 Hz), 3.04 (1H, d, J=11 Hz), 6.23 (1H, dd, J=18, 11 Hz), 7.01 (1H, d, J=8 Hz), 7.01 (1H, s), 7.10 (1H, d, J=8 Hz), 8.73 (1H, br s, indole NH)

ACKNOWLEDGMENT

Authors' heartiest thanks are due to Professor Shin-ichiro Sakai and Mr Hirokazu Maeno of Chiba University for their kind help to perform final purification and identification of our synthetic lyngbyatoxin A (teleocidin A-1) and teleocidin A-2. We thank also Dr Hirota Fujiki of National Cancer Center Research Institute for assay of the biological activities. This work was supported by a Grant-in-Aid from the Ministry of Education, Science and Culture, which is gratefully acknowledged.

REFERENCES AND NOTES

- 1 Part I Muratake, H, Natsume, M *Tetrahedron* preceding paper
- 2 Kozikowski, A P, Ishida, H *Heterocycles* **1980**, *14*, 55-58
- 3 Heinzelman, R V, Anthony, W C, Lyttle, D A, Szmuszkovicz, J *J Org Chem* **1960**, *25*, 1548-1558
- 4 Somei, M, Karasawa, Y, Kaneko, C *Heterocycles* **1981**, *16*, 941-949
- 5 (a) Corey, E J, Andersen, N H, Carlson, R M, Paust, J, Vedejs, E, Vlattas, I, Winter, R E *J Am Chem Soc* **1968**, *90*, 3245-3247 (b) McDonald, E, Martin, R T *Tetrahedron Lett* **1977**, 1317-1320
- 6 Satoh, T, Suzuki, S, Suzuki, Y, Miyaji, Y, Imai, Z *Tetrahedron Lett* **1969**, 4555-4558
- 7 Gilchrist, T L, Lingham, D A, Roberts, T G *J Chem Soc, Chem Commun* **1979**, 1089-1090
- 8 Drinkwater, D J, Smith, P W G *J Chem Soc (C)* **1971**, 1305-1307
- 9 Seki, H, Koga, K, Matsuo, H, Ohki, S, Matsuo, I, Yamada, S *Chem Pharm Bull* **1965**, *13*, 995-1000
- 10 de Laszlo, S E, Ley, S V, Porter, R A *J Chem Soc, Chem Commun* **1986**, 344-346
- 11 Shioiri, T, Ninomiya, K, Yamada, S *J Am Chem Soc* **1972**, *94*, 6203-6205
- 12 Kitagawa, Y, Oshima, K, Yamamoto, H, Nozaki, H *Tetrahedron Lett* **1975**, 1859-1862
- 13 Hamada, Y, Shioiri, T *Chem Pharm Bull* **1982**, *30*, 1921-1924
- 14 Yamada, S, Kasai, Y, Shioiri, T *Tetrahedron Lett* **1973**, 1595-1598
- 15 Nishiwaki, S, Fujiki, H, Yoshizawa, S, Suganuma, M, Furuya-Suguri, H, Okabe, S, Nakayasu, M., Okabe, K, Muratake, H, Natsume, M, Umezawa, K, Sakai, S, Sugimura, T *Jpn J Cancer Res* **1991**, *82*, 779-783
- 16 Okabe, K, Muratake, H, Natsume, M *Tetrahedron* **1990**, *46*, 5113-5120